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Attorney Docket No.: 15966-601
(CURA-101)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANTS: Gould-Rothberg *et al.*
ASSIGNEE: Curagen Corporation
SERIAL NUMBER: Filed herewith EXAMINER: Not yet assigned
FILING DATE: Filed herewith ART UNIT: Not yet assigned
FOR: METHOD OF IDENTIFYING TOXIC AGENTS USING NSAID-INDUCED
DIFFERENTIAL GENE EXPRESSION IN LIVER

JC825 U.S. PTO
09/11/321
11/20/00

November 20, 2000
Boston, Massachusetts

Assistant Commissioner for Patents
Washington, D.C. 20231

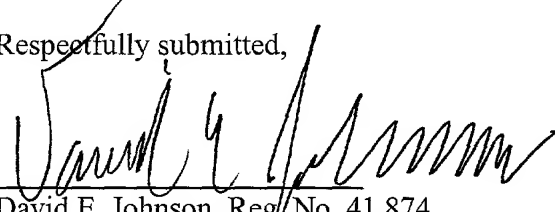
TRANSMITTAL LETTER

Transmitted herewith for filing in the present application are the following documents:

- ☒ Request for Filing a New Nonprovisional Application 37 C.F.R. §1.53(b) (2 pgs.);
- ☒ Non-provisional Patent Application (82 pgs.);
- ☒ Combined Declaration and Power of Attorney, Unexecuted (4 pgs.);
- ☒ Statement Claiming Small Entity Status, Unexecuted (2 pgs.);
- ☒ Check #7265 in the amount of \$799.00; and
- ☒ Return Postcard.

The Commissioner is hereby authorized to credit any overpayment or charge any deficiencies to Deposit Account No. 50-0311, Reference No. 15966-101. A duplicate copy of this Transmittal Letter is enclosed. If the enclosed papers are considered incomplete, the Mail Room is respectfully requested to contact the undersigned collect at (617) 542-6000, Boston, Massachusetts.

Respectfully submitted,


David E. Johnson, Reg. No. 41,874
James Gregory Cullen, Reg. No. 43,569
Attorneys for Applicants
c/o MINTZ, LEVIN
One Financial Center
Boston, Massachusetts 02111
Tel: (617) 542-6000
Fax: (617) 542-2241

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

FIRST-NAMED INVENTOR OR Gould-Rothberg
APPLICATION IDENTIFIER:

FOR: METHOD OF IDENTIFYING TOXIC AGENTS USING NSAID-INDUCED
 DIFFERENTIAL GENE EXPRESSION IN LIVER

November 20, 2000
Boston, Massachusetts

Box PATENT APPLICATION
Assistant Commissioner for Patents
Washington, D.C. 20231

**REQUEST FOR FILING A NEW NONPROVISIONAL APPLICATION
UNDER 37 C.F.R. §1.53(b)**

1. This is a request for filing a new nonprovisional application under 37 C.F.R. §1.53(b).
2. ☒ Specification and Drawings (Total pages: 82);
 Specification (74 pages); Claims (7 pages); Abstract (1 page); and
 Drawings: 0 sheets; FIGS. _____.
 ☐ Formal
 ☐ Informal
3. ☒ Declaration and Power of Attorney
 ☒ Unsigned
 ☐ Signed
4. ☐ Information Disclosure Statement (IDS)
 ☐ Copy of IDS and PTO-1449 (___ pages)
 ☐ Copies of references cited
5. ☐ Assignment Papers
 ☐ Recordation Form Cover Sheet (PTO-1595)
 ☐ Assignment Document
6. ☐ Statement Claiming Small Entity Status
 ☐ Claiming Small Entity As Independent Inventor (37 C.F.R. §§1.9(f) & 1.27(b)).
 ☒ Claiming Small Entity As Small Business Concern (37 C.F.R. §§1.9(f) & 1.27(c)).
 ☐ Claiming Small Entity As Nonprofit Organization (37 C.F.R. §§1.9(f) & 1.27(d)).

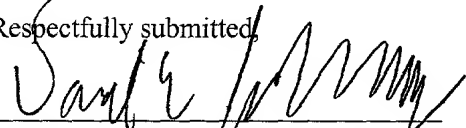
FIRST-NAMED INVENTOR OR
APPLICATION IDENTIFIER: Gould-Rothberg
Request for New Nonprovisional Application (37 C.F.R. §1.53(b))

7. Fee Calculation

CLAIMS AS FILED					
Claims	Number Filed	Basic Fee Allowance	Number Extra	Rate	Basic Fee 37 C.F.R. 1.16(a) \$710.00
Total Claims (37 C.F.R. 1.16(c))	56	- 20 =	36	\$ 18.00	648.00
Independent Claims (37 C.F.R. 1.16(b))	6	- 3 =	3	\$80.00	240.00
Multiple Dependent Claim(s), if any (37 C.F.R. 1.16(d))	0			\$270.00	0
SUBTOTAL:					\$1598.00
Reduction by 50% for filing by small entity:					- \$799.00
TOTAL FEE:					\$799.00

8. ☒ A check # 7265 in the amount of \$799.00 is enclosed.
9. ☒ The Commissioner is hereby authorized to credit overpayments or charge the following fees to Deposit Account No. 50-0311, Ref. No. 15966-101:
- ☒ Fees required under 37 C.F.R. §1.16;
- ☒ Fees required under 37 C.F.R. §1.17;
- ☒ Fees required under 37 C.F.R. §1.18.
10. ☒ Return Receipt Postcard Enclosed.
11. ☐ Other Documents Enclosed:
- ☐ Change of Attorney Address In Application.
- ☐ Limited Recognition under 37 C.F. § 10.9(b) for Michel Morency.

Respectfully submitted,


David E. Johnson, Reg. No. 41,874
James Gregory Cullem, Reg. No. 43,569
Attorney(s) for Applicants
MINTZ, LEVIN, COHN, FERRIS,
GLOVSKY and POPEO, P.C.
One Financial Center
Boston, Massachusetts 02111
Tel: (617) 542-6000
Fax: (617) 542-2241

Dated: NOVEMBER 20, 2000

TRADOCS:1402288.1(%20G01!.DOC)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANTS: Gould-Rothberg *et al.*
ASSIGNEE: Curagen Corporation
SERIAL NUMBER: Filed herewith EXAMINER: Not yet assigned
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**STATEMENT CLAIMING SMALL ENTITY STATUS
(37 CFR 1.9(f) and 1.27(c) – SMALL BUSINESS CONCERN)**

I hereby state that I am an official of the small business concern empowered to act on behalf of the concern identified below:

**CURAGEN CORPORATION
322 East Main Street
Branford, CT 06405**

I hereby state that the above identified small business concern qualifies as a small business concern, as defined in 13 C.F.R. § 121, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees to the United States Patent and Trademark Office under Sections 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third-party or parties controls or has the power to control both.

I hereby state that rights under contract or law have been conveyed to, and remain with, the small business concern identified above with regard to the invention described in:

- ☐ the specification filed herewith with title as listed above;
☒ the application identified above; or
☐ the patent identified above.

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organization having rights in the invention is listed below and no rights to the invention are held by any person, other than the inventor, who would not qualify as an independent inventor under 37 C.F.R. § 1.9(c), if that person made the invention, or by any concern which would not qualify as a small business concern under 37 C.F.R. § 1.9(d) or a nonprofit organization under

APPLICANTS: Gould-Rothberg *et al.*
U.S.S.N.: Filed herewith

37 C.F.R. § 1.9(e).

Each person, concern, or organization having any rights in the invention is listed below:

- ☒ no such person, concern, or organization exists.
☐ each such person, concern, or organization is listed below: ♦
♦ *Note: Separate Statements Claiming Small Entity Status are required from each named person, concern, or organization having rights to the invention. 37 C.F.R. § 1.27.*

Full Name:

Address:

☐ Individual ☐ Small Business Concern ☐ Non-profit Organization

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small business entity is no longer appropriate. 37 C.F.R. § 1.28(b).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

Name of person signing: **George Xixis, Esq.**

Title in organization of person signing: Patent Counsel

Address of person signing: CURAGEN CORPORATION
322 East Main Street
Branford, CT 06405

Signature: _____

Date: _____

TRADOCS:1402293.1(%20L01!.DOC)

METHOD OF IDENTIFYING TOXIC AGENTS USING NSAID-INDUCED DIFFERENTIAL GENE EXPRESSION IN LIVER

RELATED APPLICATIONS

This application claims priority to USSN 60/166,923, filed November 22, 1999, and
5 USSN 60/183,531, filed February 18, 2000, the disclosures of which are hereby incorporated by
reference herein.

FIELD OF THE INVENTION

The invention relates generally to nucleic acids and polypeptides, and more particularly
to the identification of differentially expressed nucleic acids and proteins in liver.

BACKGROUND OF THE INVENTION

10 Liver is the primary organ for biotransformation of chemical compounds and their
detoxification. Liver injury produced by chemicals has been recognized for over 100 years, and
hepatic damage is one of the most common toxicities among drugs at pre-clinical and clinical
stages of drug development. Over 30% of new chemical entities (NCE) are generally terminated
15 due to adverse liver effects in humans. During a period of 30 years, hepatotoxicity has been the
major cause of drug withdrawal for safety reasons at the marketing stage, accounting for 18%
overall drug withdrawal. Many of the drugs that are withdrawn from market due to
hepatotoxicity produce lethality in a small percentage of patient population and are classified as
type II lesion (or idiosyncratic, sporadic) toxicity.

20 Non-steroidal anti-inflammatory drugs (NSAIDs) are a group of unrelated chemical
compounds that have been used to successfully treat rheumatic and musculoskeletal disease.
Unfortunately, unwanted hepatotoxic side effects have led to the premature market withdrawal
of several NSAIDs, including Cincophen, Benoxaprofen, Piroxicam, Suprofen, and Bromfenac.
The pervasiveness of idiosyncratic reactions of many NSAIDs has led the Food and Drug
25 Administrations Arthritis Advisory Board to conclude that NSAIDs as a group should be
considered to induce hepatotoxicity.

It is estimated that annual NSAIDS consumption in the U.S exceeds 10,000 tons. Due to this large consumption of NSAIDS for a wide variety of pain and inflammatory conditions, it has become an important class of drugs responsible for liver injury, despite the overall extremely low incidence of producing hepato-toxicity. Liver injury resulting from NSAIDs can have several forms, including acute toxicity resulting from hepatocellular (parenchymal) damage (*e.g.* necrosis) and arrested bile flow (cholestasis). The general mechanism that is thought to mediate NSAIDS toxicity is idiosyncratic reaction (Type II) to the drug (both immunologic and metabolic), which is dose independent, and presumably results from interindividual variation in drug metabolism. Currently no clear mechanism of drug-induced idiosyncratic toxicity is available. Accordingly, there remains a great need to elucidate the molecular basis of idiosyncratic hepatotoxicity, such as NSAID-induced toxicity, including the identification of genes and proteins differentially expressed in response to administration of such drugs.

SUMMARY OF THE INVENTION

In accordance with the present invention, there are provided methods of screening and identifying test agents which induce hepatotoxicity, *e.g.* idiosyncratic hepatotoxicity. The methods of the invention are based in part on the discovery that certain nucleic acids are differentially expressed in liver tissue of animals treated with NSAIDs. These differentially expressed nucleic acids include novel sequences that, while previously described, have not heretofore been identified as responsive to drugs, such as NSAIDs, which induce idiosyncratic hepatotoxicity.

In various aspects, the invention includes a method of screening a test agent for toxicity, *e.g.*, idiosyncratic hepatotoxicity. For example, in one aspect, the invention provides a method of identifying a hepatotoxic agent by providing a test cell population comprising a cell capable of expressing one or more nucleic acids sequences responsive to drugs, *e.g.* NSAIDs, which induce idiosyncratic hepatotoxicity, contacting the test cell population with the test agent and comparing the expression of the nucleic acids sequences in the test cell population to the expression of the nucleic acids sequences in a reference cell population. An alteration in expression of the nucleic acids sequences in the test cell population compared to the expression of the gene in the reference cell population indicates that the test agent is hepatotoxic. In one aspect, expression in

the test cell population is compared to the expression of a reference cell population exposed to a NSAID that is classified as low risk, very low risk, or overdose risk of hepatotoxicity, thereby to predict whether the test agent has low, very low, or overdose risk of hepatotoxicity. In another aspect, the test cell population is compared to the expression of a reference cell population exposed to a NSAID which induces a known type of hepatic injury, *e.g.* hepatocellular damage, cholestasis, or elevated transaminase level, thereby to predict whether the test agent is likely to induce a given type of hepatotoxic injury.

In a further aspect, the invention provides a method of assessing the hepatotoxicity, *e.g.* idiosyncratic hepatotoxicity, of a test agent in a subject. The method includes providing from the subject a cell population comprising a cell capable of expressing one or more NSAID-responsive genes, and comparing the expression of the nucleic acids sequences to the expression of the nucleic acids sequences in a reference cell population that includes cells from a subject whose exposure status to a hepatotoxic agent is known. An alteration in expression of the in the test cell population compared to the expression of the nucleic acids sequences in the reference cell population indicates the hepatotoxicity of the test agent in the subject.

Also provided are novel nucleic acids, as well as their encoded polypeptides, whose expression is responsive to the effects of NSAIDS.

Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In the case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

DETAILED DESCRIPTION OF THE INVENTION

The invention is based in part on the discovery of nucleic acid sequences which are differentially expressed in rodent liver cells upon administration of NSAIDS. The discovery includes groups of nucleic acid sequences whose expression is correlated with hepatotoxicity risk associated with, and injury type induced by, NSAID administration.

The differentially expressed nucleic acid sequences were identified by examining 29 different NSAIDS that have varying degrees of hepatotoxicity. These 29 drugs, shown in Table 1, below, were first categorized as low dose (1-75 mg/kg) and high dose (above 75 mg/kg) drugs, then classified as non-toxic, toxic, and those withdrawn from market (within each dose). Each of the 29 NSAIDS was administered orally to groups (3 animals per group) of 12 week old male Sprague Dawley rats for 72 hours (3 days) at the dosages specified in Table 1 (e.g. Naproxen: 54 mg/kg/day PO in QD x 3 days in H₂O). Vehicle controls (water, ethanol, canola oil) were also included (3 animals per group). The animals were sacrificed 24 hours after the final dose, liver tissue was removed on necroscopy, and total RNA was recovered from the dissected tissue. Complementary DNA (cDNA) was prepared and samples were processed through GENECALLING™ differential expression analysis, as described in U.S. Patent No. 5,871,697 and in Shimkets *et al.*, *Nature Biotechnology* 17: 798-803 (1999), the disclosures of which are hereby incorporated by reference herein.

Table 1: NSAIDS and Dosages Administered

Compound	Dose	Vehicle
<i>Acetaminophen</i>	133 mg/kg/day p.o. in QD x 3 days.	10 %Ethanol Vehicle
<i>Acetylsalicylic Acid</i>	200 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Benoxaprofen</i>	16 mg/kg/day p.o. in QD x 3 days.	H2O Vehicle
<i>Bromfenac</i>	7.5 mg/kg/day p.o. in QD x 3 days.	H2O Vehicle
<i>Celecoxib</i>	89 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Diclofenac</i>	38 mg/kg/day p.o. in QD x 3 days.	H2O Vehicle
<i>Etodolac</i>	30 mg/kg/day p.o. in QD x 3 days.	10% Ethanol Vehicle
<i>Felbinac</i>	33 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Fenoprofen</i>	154 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Flurbiprofen</i>	10 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Ibuprofen</i>	211 mg/kg/day p.o. in QD x 3 days.	H2O Vehicle
<i>Indomethacin</i>	4 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Ketoprofen</i>	10 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Ketorolac</i>	1.5 mg/kg/day p.o. in QD x 3 days.	10% Ethanol Vehicle
<i>Meclofenamate</i>	20 mg/kg/day p.o.in QD x 3 days.	H2O Vehicle

<i>Mefenamic Acid</i>	79 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Nabumetone</i>	143 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Naproxen</i>	54 mg/kg/day p.o. in QD x 3 days.	10% Ethanol Vehicle
<i>Olsalazine</i>	222 mg/kg/day p.o. in QD x 3 days.	H2O Vehicle
<i>Oxaprozin</i>	100 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Phenacetin</i>	100 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Phenylbutazone</i>	100 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Piroxicam</i>	20 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Sulindac</i>	77 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Sulphasalazine</i>	338 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Suprofen</i>	20 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Tenoxicam</i>	10 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle
<i>Tolmentin</i>	100 mg/kg/day p.o. in QD x 3 days.	H2O Vehicle
<i>Zomepirac</i>	19 mg/kg/day p.o. in QD x 3 days.	Canola Oil Vehicle

3635 gene fragments were initially found to be differentially expressed in rat liver tissue (analysis of variance, $p < 0.01$) in response to these compounds. The compounds were then classified according to hepatotoxicity risk, as indicated in Table 2.

Table 2: Hepatotoxicity Risk of NSAIDs

Compound	Risk
Acetaminophen	Overdose Risk
Acetylsalicylic Acid	Overdose Risk
Benoxaprofen	Low Risk
Bromfenac	Low Risk
Celecoxib	Unknown
Diclofenac	Low Risk
Etodolac	Very Low Risk
Felbinac	Unknown
Fenoprofen	Very Low Risk
Flurbiprofen	Very Low Risk
Ibuprofen	Very Low Risk
Indomethacin	Very Low Risk
Ketoprofen	Very Low Risk
Ketorolac	Unknown

Meclofenamate	Very Low Risk
Mefenamic Acid	Very Low Risk
Nabumetone	Very Low Risk
Naproxen	Very Low Risk
Olsalazine	Unknown
Oxaprozin	Very Low Risk
Phenacetin	Overdose Risk
Phenylbutazone	Low Risk
Piroxicam	Very Low Risk
Sulindac	Low Risk
Sulphasalazine	Unknown
Suprofen	Very Low Risk
Tenoxicam	Very Low Risk
Tolmentin	Very Low Risk
Zomepirac	Very Low Risk

In order to discriminate among these groups, the above compound set was divided into a training set (consisting of three compounds per group), and a test set (consisting of the remainder. This was done to minimize the reliance on the assumptions required for parametric analyses. Compounds with unknown risk were not used in this analysis. The training set employed is shown in Table 3.

Table 3: Training Set of NSAIDs by Risk Classification

Control	Low Risk	Very Low Risk	Overdose Risk
Sterile water	Benoxaprofen	Flurbiprofen	Acetaminophen
10% Ethanol	Phenylbutazone	Oxaprozin	Acetylsalicylic Acid
Canola oil	Sulindac	Tenoxicam	Phenacetin

The 3635 differentially expressed nucleic acid fragments were then analyzed using a stepwise multivariate analysis of variance as follows:

1. Calculate 3635 T2 (yi1) (Hoettelling's trace, one of the test statistics used for this analysis) values, one for each differentially expressed fragment. The fragment with the largest individual T2 value is selected as the first discriminatory set (yi1).
2. Calculate 3634 T2 (yi1,yi2) values, one for each combination of two fragments. The fragment pair with the largest individual T2 value are selected as the second discriminatory set.

3. Calculate 3626 T2 ($y_{i1}, y_{i2}, y_{i3}, \dots, y_{i,10}$), one for each combination of ten fragments.

The fragment set with the largest T2 value are selected as the final discriminatory set.

This stepwise procedure is used whenever the number of dependent variables (gene fragments) exceeds the number of independent variables (samples). In addition to fragment addition, fragment elimination occurs whenever an added fragment no longer contributes significant discriminatory power to the existing set. This eliminates bias as to the order fragments enter the model (Ahrens and Lauter, Mehrdimensionale Varianzanalyse, Akademie-Verlag, Berlin (1974); Dziuda, *Medical Inform.* 15(4): 319 (1990)).

This analysis protocol identified ten fragments that significantly ($p=6.02 \times 10^{-28}$) discriminated among the drugs in the test set. Two fragments on this list were not required to maintain the discriminatory ability and were subsequently removed ($p=3.96 \times 10^{-26}$). Differential expression of these gene fragments were successfully confirmed using an unlabeled oligonucleotide competition assay (Shimkets *et al.*, *Nature Biotechnology* 17: 198-803 (1999)). The 8 fragments (RISKMARKER 1-8) represent both novel and known rat genes for which the sequence identity to genes in public databases is either high ($>90\%$), moderate (70-90%), or low ($<70\%$).

The identity of these 8 hepatocitiy risk discriminatory nucleic acid sequences(with GenBank accession numbers) are further described below. Where appropriate, the cloned sequence from isolation is provided; this sequence was then extended using either Genbank rat ESTs or from internally (Curagen Corporation) sequenced rat fragments. The extended contig sequence is provided as "consensus." Finally, the best BlastN and BlastX results are also provided. In some instances the cloned sequence is identical to a known rat gene, in those instances the name of the gene and a database accession number and the sequence listed in the database is provided:

RISKMARKER 1

RISKMARKER 1 is a novel 1265 bp gene fragment , which has 67% sequence identity to human rac1 genomic fragment [AJ132695], probable 3' UTR. The nucleic acid was initially identified in a cloned fragment having the following sequence:

1 caattgaaaa aagtttggttc tagtggtcga aaggcccaac actgtgttct tgccagtga

55

Identified = 270/354 (76%); Positives = 270/354 (76%); Strand = Minus / Plus

Query: 357 TGAAACATCACTGTCT-TGCCAGAGTACCAACTGTCATGTGATTGATGCCGCCCCCTC 299
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Sbjct: 27795 TGCCAGATTACCAGACTGTCACTTGACCAATACTGACCC-TCCTTT-ACCTCGCCCACGC 27852

60

Query: 298 TAGAC-CTCACCCAC-GCGGACACAT-GCTTCC-G--GTACCTTTGGGTCTGTGAGGTTT 245
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Sbjct: 27853 G-GACACCGCCTCTCT GT|GTCGCTTTGCCTATTGATGTTCTCTTTGGGTCTGTGAGGTTT 27911

65

Query: 244 TGTCAA--GCGCTAGTGCTAACGCCGTTCTGTACAACCTAACTCACTGGCAAGAACACAG 187
TGT AA G |TAGTG|T A|G GTT|TGTA|AA| TAA|T|A|TGG| AGAA A|AG
Sbjct: 27912 TGTAACCTGTGTAGTGTGACGATGTTCTGTACAACCTAACTCACTGGCGAGAATACAG 27971

Query: 186 TGT'TGGG'CCTTT'CGACCACTAGAACAACTTTTTCAATTGACAGTTGCAGAATTGTGGGA 127
GT GG || TT| ||A|TA AA| A TTTT'AATTGA|AGTTG|AGAATTGTGGGA
70 Sbjct: 27972 CGTGGGACCCTT'CAGCCACTACAACAGAATTTTTTAAATTGACAGTTGCAGAATTGTGGGA 28031

Query: 126 GTGTTTTTACATTGATCTTTTGCTAATGCAGTTAGCAGTATGTTTTGCATGTATGACTTA 67
GTGTTTTTA|ATTGAT|TTTTG|TAATG|A TTAG|A TATGTTTG|ATGTATGA|TTA
Sbjct: 28032 GTGTTTTTACATTGATCTTTTGCTAATGCAATTAGCATTATGTTTTGCATGTATGACTTA 28091

5 Query: 66 ATAAATCCTTGAATCATAAAAAAAAAAAAAAAAAATGCTTTGGAACTTGAAAAAAAA 10
ATAAAT||TTGAAT|ATA A AA A TGT TTTG A|TTGA AA AA
Sbjct: 28092 ATAAATCCTTGAATCATAACGACTGGTAATACTGGTGTTTTGTGAGACTTGATGAACAA 28148

RISKMARKER2

10 RISKMARKER2 is a 650 bp rat expressed sequence tag (EST) [AW435096]. The nucleic acid sequence was initially identified in a cloned fragment having the following sequence:

1 TTTTTTTTTTTTTTTTTTGGCAGAATTCTGATGTTTACTGGGACCCATAGTAGTCAAGGTGACAGCAAGGGTAGGGGAGGA
81 AACTCAGCAGAGGCGGATCCAGGTCTGGAGGGAAGCTGACAGCAGCCCAGTAAGCTGTGCCAGAAGGCTGTAACAGTAG
161 CGGAGCCAGTGACAGCGCCAGGCTGGGCTGGGTTCTCTCTGTGGGTGTGCACGGCAAAGCTGCGGCCTGTGGGCCCTGGG
241 GGGCCTGTGAGCTCCACATCCACCACATGCATGTCGGTGAGGCTAAGGTGAGCCACAAGCACCCCAATGACACGATCAAA
321 GCCTAGACTGGGAGCGGCCAGGGCAGCGCTGCCATGGTGTGGAGTTTCGGGGGGCCAAGGGGCAGAGCCCACGCACAG
401 GGCCCTCATAGAGCACTGTGCGGGGCCACTACTATGTGCGGCAGCCAGGGGTCNCTCCAGCCGGAAGCCATCAGGATGT
481 GTGG (SEQ ID NO: 3)

15 The cloned sequence was assembled into a contig resulting in the following consensus sequence:

1 TTTTTTTTTTTTTTTTTTGGCAGAATTCTGATGTTTACTGGGACCCATAGTAGTCAAGGTGACAGCAAGGGTAGGGGAGGA
81 AACTCAGCAGAGGCGGATCCAGGTCTGGAGGGAAGCTGACAGCAGCCCAGTAAGCTGTGCCAGAAGGCTGTAACAGTAG
161 CGGAGCCAGTGACAGCGCCAGGCTGGGCTGGGTTCTCTCTGTGGGTGTGCACGGCAAAGCTGCGGCCTGTGGGCCCTGGG
241 GGGCCTGTGAGCTCCACATCCACCACATGCATGTCGGTGAGGCTAAGGTGAGCCACAAGCACCCCAATGACACGATCAAA
321 GCCTAGACTGGGAGCGGCCAGGGCAGCGCTGCCATGGTGTGGAGTTTCGGGGGGCCAAGGGGCAGAGCCCACGCACAG
401 GGCCCTCATAGAGCACTGTGCGGGGCCACTACTATGTGCGGCAGCCAGGGGTCCTCCAGCCGGAAGCCATCAGGATGT
481 GTGGCCATGGTGACTCGAAGGCTCTGGAGGCCTCCGGCTGCATCCAATCTGCTGATGTCTTCACAACCCACAGGGCCCC
561 TCGGGCCACAAACACCGTGTGGCCCCAGTGGTTTGAAGCCTCCAGGAGCTGCCGCTCTGTGGTCTGGTCAGCGAGAGCTG
641 AGGGGGATCC (SEQ ID NO: 4)

Blast-N Results:

20 >gb:GENBANK-ID:AW435096|acc:AW435096 UI-R-BJ0p-afy-e-10-0-UI.s1 UI-R-BJ0p
Rattus norvegicus cDNA clone UI-R-BJ0p-afy-e-10-0-UI 3', mRNA sequence -
Rattus norvegicus, 484 bp (RNA).
Length = 484

Plus Strand HSPs:

Score = 2413 (362.0 bits), Expect = 1.2e-102, P = 1.2e-102
Identities = 483/484 (99%), Positives = 483/484 (99%), Strand = Plus / Plus

5 Query: 1 TTTTTTTTTTTTTTTTGGCAGAAATCTGATGTTTACTGGGACCCATAGTAGTCAAGGTG 60
Sbjct: 1 TTTTTTTTTTTTTTTTGGCAGAAATCTGATGTTTACTGGGACCCATAGTAGTCAAGGTG 60

10 Query: 61 ACAGCAAGGGTAGGGGAGGAACTCAGCAGAGGCGGATCCCAGGTCTGGAGGGAAGCTGA 120
Sbjct: 61 ACAGCAAGGGTAGGGGAGGAACTCAGCAGAGGCGGATCCCAGGTCTGGAGGGAAGCTGA 120

15 Query: 121 CAGCAGCCAGTAAGCTGTGCCAGAAGGCTGTAACAGTAGCGGAGCCAGTGACAGCGCCA 180
Sbjct: 121 CAGCAGCCAGTAAGCTGTGCCAGAAGGCTGTAACAGTAGCGGAGCCAGTGACAGCGCCA 180

20 Query: 181 GGCTGGGCTGGGTTCTCTCTGTGGGTGTGCACGGCAAAGCTGCGGCCTGTGGGCCCTGGG 240
Sbjct: 181 GGCTGGGCTGGGTTCTCTCTGTGGGTGTGCACGGCAAAGCTGCGGCCTGTGGGCCCTGGG 240

25 Query: 241 GGGCCTGTCAGCTCCACATCCACCACATGCATGTCGGTGAGGCTAAGGTCAGCCACAAGC 300
Sbjct: 241 GGGCCTGTCAGCTCCACATCCACCACATGCATGTCGGTGAGGCTAAGGTCAGCCACAAGC 300

30 Query: 301 ACCCCAATGACACGATCAAGCCCTAGACTGGGAGCGGCCAGGGCAGCGGCTGCCATGGTG 360
Sbjct: 301 ACCCCAATGACACGATCAAGCCCTAGACTGGGAGCGGCCAGGGCAGCGGCTGCCATGGTG 360

35 Query: 361 TTGGAGTTTCGGGGGGCCAAAGGGGAGAGCCACGCACAGGGCCCTCATAGAGCACTGTG 420
Sbjct: 361 TTGGAGTTTCGGGGGGCCAAAGGGGAGAGCCACGCACAGGGCCCTCATAGAGCACTGTG 420

40 Query: 421 CGGGGCCCCACTACTATGTGCGGCAGCCAGGGGTCCCTCCAGCCGGAAGCCATCAGGATGT 480
Sbjct: 421 CGGGGCCCCACTACTATGTGCGGCAGCCAGGGGTCCCTCCAGCCGGAAGCCATCAGGATGT 480

Query: 481 GTGG 484
Sbjct: 481 GTGG 484

Blast-X Results:

>ptnr:SPTREMBL-ACC:Q19527 F17C8.3 PROTEIN - Caenorhabditis elegans, 973 aa.

[Top](#) [Previous Match](#) [Next Match](#)

Length = 973

Minus Strand HSPs:

Score = 351 (123.6 bits), Expect = 6.3e-30, P = 6.3e-30
Identities = 78/161 (48%), Positives = 96/161 (59%), Frame = -1

Query: 650 GSPSALADQTTTERQLLEASNHWGHTVFVARGALWGCEDISRLDAAGGLQSLRVMTATHPD 471
GSP+ A+Q +L+S G++ GALWG DI++ GL+LVTM HP
Sbjct: 530 GSPTCFANQELLEKLTKLSLSHGKKLLIPAGALWGANDIQKMADVGSLSKGLTVTMIKHPT 589

Query: 470 GFRLEGPLAAAHSSGP----RTVLYEGPVRGLCPLAPRNSNTMAAAALAAPSLGFDRVI 306
F+L PL + TVLYEG VRGLCPLAP N NTMA ALAA +LGFD V
Sbjct: 590 SFKLGSPLEINEKAKLEETNETVLYEGSVRGLCPLAPNNVNTMAGGALAASNLGFDEVK 649

Query: 305 GVLVADLSLTDHVVVDVELTGPPGPTGRSFAVHTHRENPAPGAVT 168
L++D +TD HVV+V+G G F V T R NPA+PGAVT
Sbjct: 650 AKLISDPKMTDWHVVEVRVEGDDG----FEVITRRNNPAKPGAVT 690

RISKMARKER3

RISKMARKER3 is a 1019 nucleotide sequence encoding superoxide dismutase copper chaperone [AF255305]:

```

1      ggtctctgga ccctaccggt tgtgtggccc aagcgggtga ctgcagccag gatggcttcg
61     aagtcggggg acggtggaac tatgtgtgcg ttggagttta cagtacagat gagttgtcag
5      121     agctgcgtgg acgctgtgca caagaccctg aaaggggagg cgggtgtcca gaatgtggaa
181    gttcagttgg agaaccagat ggtgttgggt cagaccactt tgcccagcca ggaggtgcaa
241    gcgctcctgg aaagcacagg gaggcaggct gtactcaagg gcatgggcag cagccaacta
301    aagaatctgg gagcagcagt ggccattatg gagggcagtg gcaccgtaca gggggtgggtc
361    cgcttcctac agctgtcctc tgagctctgc ctgattgagg gaaccatcga cggcctggag
10     421    cctgggctgc atgggcttca tgtccatcag tatggggacc ttaccaagga ctgcagcagc
481    tgtggggacc attttaaccc tgatggagca tctcatgggg gtccctcagga cactgatcgg
541    caccggggag atctgggcaa tgttcacgct gaagctagtg gccgagctac cttccggata
601    gaggataaac agctgaaggt gtgggatgtg attggccgca gtctggttgt tgatgagggg
661    gaagatgacc tgggcccggg aggccatccc ttatccaagg tcacagggaa ttctgggaag
15     721    aggttggcct gtggcatcat tgcacgctct gctggccttt tccagaatcc caagcagatc
781    tgctcctgtg atgggctcac tatctgggag gagcgaggcc ggccattgct tggccaaggc
841    cgaaaggact cagcccaacc cctgtctcac ctctgaacag agcctcctgt caggttattc
901    agtctccta gctgaacatc ttctgcaga gggagcctca agcccttgct tgtataggcc
961    taaagggcag ataggcattg ttgtatcctg agcaaattaa attgttactc tcatatggc

```

RISKMARKER4

RISKMARKER4 is a 878 nucleotide sequence encoding alpha-2 microglobulin [U31287]:

```

1      ggcacgagca gagagattgt cccaacagag aggcaattct attccctacc aacatgaagc
25     61     tgttgctgct gctgctgtgt ctgggcctga cactggtctg tggccatgca gaagaagcta
121    gttccacaag agggaacctc gatgtggcta agctcaatgg ggattggttt tctattgtcg
181    tggcctctaa caaaagagaa aagatagaag agaattggcag catgagagtt tttatgcagc
241    acatcgatgt cttggagaat tccttaggct tcaagttccg tattaaggaa aatggagagt
301    gcaggggaact atatttgggt gcctacaaaa cgccagagga tggcgaatat tttgttgagt
30     361    atgacggagg gaatacattt actatactta agacagacta tgacagatat gtcatgtttc
421    atctcattaa tttcaagaac ggggaaacct tccagctgat ggtgctctac ggcagaacaa
481    aggatctgag ttcagacatc aaggaaaagt ttgcaaaact atgtgaggcg catggaatca
541    ctagggacaa tatcattgat ctaaccaaga ctgacgctg tctccaggcc cgaggatgaa
601    gaaaggcctg agcctccagt gctgagtgga gacttctcac caggactcta gcatcaccat
35     661    ttctgttcca tggagcatcc tgagacaaat tctgcatctt gatttccatc ctctgtcaca
721    gaaaagtgca atcctgggtc ctccagcatc ttccctaggt taccaggagc aacacatcga
781    gaattaaaag ctttcttaaa tttctcttgg cccaccccat gatcattccg cacaaatc
841    ttgctcttgc agttcaataa atgattaccc ttgcaactt

```

RISKMARKER5

RISKMARKER5 is a 2443 bp rat mRNA for Mx3 protein [X52713]. The nucleic acid was initially identified in a cloned fragment (having 100% sequence identity to the rat mRNA) having the following sequence:

```

1 CCATGGATGAAATCTTCCAGCATCTGAATGCCTACCGCCAGGAGGCTCACAACCTGCATCTCCAGCCACATTCCATTGATC
81 ATCCAGTATTTTCATCTTGAAGATGTTTGCTGAGAAGCTGCAGAAGGGCATGTCCAGCTCCTGCAGGACAAGGATTCCTG
161 CAGCTGGCTCCTGAAGGAAAAGAGTGACACCAAGTGAGAAGAGGAGATTCTGAAGGAGCGGTTGGCAAGGCTGGCCCAAG
241 CTCAGCGCAGGCTAGC (SEQ ID NO: 5)

```

Blast-N Results:

>gb:GENBANK-ID:RNMX3|acc:X52713 Rat mRNA for Mx3 protein - Rattus norvegicus, 2443 bp.

Top Previous Match Next Match
Length = 2443

Plus Strand HSPs:

Score = 1280 (192.1 bits), Expect = 9.5e-52, P = 9.5e-52
Identities = 256/256 (100%), Positives = 256/256 (100%), Strand = Plus / Plus

```

Query:      1 CCATGGATGAAATCTTCCAGCATCTGAATGCCTACCGCCAGGAGGCTCACAACCTGCATCT 60
            |||
Sbjct:    1710 CCATGGATGAAATCTTCCAGCATCTGAATGCCTACCGCCAGGAGGCTCACAACCTGCATCT 1769

Query:     61 CCAGCCACATTCCATTGATCATCCAGTATTTTCATCTTGAAGATGTTTGCTGAGAAGCTGC 120
            |||
Sbjct:    1770 CCAGCCACATTCCATTGATCATCCAGTATTTTCATCTTGAAGATGTTTGCTGAGAAGCTGC 1829

Query:    121 AGAAGGGCATGCTCCAGCTCCTGCAGGACAAGGATTCCTGCAGCTGGCTCCTGAAGGAAA 180
            |||
Sbjct:    1830 AGAAGGGCATGCTCCAGCTCCTGCAGGACAAGGATTCCTGCAGCTGGCTCCTGAAGGAAA 1889

Query:    181 AGAGTGACACCAAGTGAGAAGAGGAGATTCTGAAGGAGCGGTTGGCAAGGCTGGCCCAAG 240
            |||
Sbjct:    1890 AGAGTGACACCAAGTGAGAAGAGGAGATTCTGAAGGAGCGGTTGGCAAGGCTGGCCCAAG 1949

Query:    241 CTCAGCGCAGGCTAGC 256
            |||
Sbjct:    1950 CTCAGCGCAGGCTAGC 1965

```

Blast-X Results:

>ptnr:SWISSPROT-ACC:P18590 INTERFERON-INDUCED GTP-BINDING PROTEIN MX3 - Rattus norvegicus (Rat), 659 aa.

Top Previous Match Next Match
Length = 659

Plus Strand HSPs:

Score = 429 (151.0 bits), Expect = 5.3e-39, P = 5.3e-39
Identities = 84/84 (100%), Positives = 84/84 (100%), Frame = +3

```

Query:      3 MDEIFQHLNAYRQEAHNCISSHIPLIIQYFILKMF AEKLQKGM LQLLQDKDSCSWLLKEK 182
            MDEIFQHLNAYRQEAHNCISSHIPLIIQYFILKMF AEKLQKGM LQLLQDKDSCSWLLKEK
Sbjct:    571 MDEIFQHLNAYRQEAHNCISSHIPLIIQYFILKMF AEKLQKGM LQLLQDKDSCSWLLKEK 630

Query:    183 SDTSEKRRFLKERLARLAQAQRRL 254
            SDTSEKRRFLKERLARLAQAQRRL
Sbjct:    631 SDTSEKRRFLKERLARLAQAQRRL 654

```

RISKMARKER6

RISKMARKER6 is 369 bp novel gene fragment, which has 98% amino acid identity (90% nucleic acid sequence identity) to Human ERj3 protein [AJ250137]. The nucleic acid sequence was initially identified in a cloned fragment having the following sequence:

```

1 TCTAGAAAGTCACCTTGAAGAAGTGTACGCAGGGAACCTTTGTGGAAGTAGTTAGAAACAAGCCCGTAGCCAGGCAGGCT
81 CCTGGCAAACGGAAATGCAACTGTCTGGCAGGAGATGCGAACCACACAGCTGGGACCAGGGCGCTTCCAAATGACCCAGGA
161 AGTGGTTTGTGACGAGTGCCTTAATGTCAAAGTGTGAATGAAGAACGAACACTAGAAGTGGAAATAGAGCCTGGGGTGA
241 GAGATGGCATGGAGTACCCCTTTATTGGAGAAGGTGAGCCTCATGTGGATGGGGAACCCGGAGACTTACGGTTCCGAATC
321 AAAGTTGTCAAGCACCGGATATTTGAGAGGAGAGGGGATGACCTGTACA (SEQ ID NO: 6)

```

Blast-N Results:

>gb:GENBANK-ID:HSA250137|acc:AJ250137 Homo sapiens mRNA for ERj3 protein (ERj3 gene) - Homo sapiens, 1159 bp.

Top Previous Match Next Match
Length = 1159

Plus Strand HSPs:

Score = 1524 (228.7 bits), Expect = 5.6e-63, P = 5.6e-63
Identities = 334/369 (90%), Positives = 334/369 (90%), Strand = Plus / Plus

```

Query:      1 TCTAGAAAGTCACCTTGAAGAAGTGTACGCAGGGAACCTTTGTGGAAGTAGTTAGAAACA 60
             TCTAGAA GTCAC TTGAAGAAGT TA GCAGG AA TTTGTGGAAGTAGTTAGAAACA
Sbjct:     431 TCTAGAA-GTCACCTTGAAGAAGTATATGCAGGAAATTTTGTGGAAGTAGTTAGAAACA 489

Query:      61 AGCCCGTAGCCAGGCAGGCTCCTGGCAAACGGAAATGCAACTGTCTGGCAGGAGATGCGAA 120
             A CC GT GC AGGCAGGCTCCTGGCAAACGGAA TGCAA TGTCGGCA GAGATGCG A
Sbjct:     490 AACCTGTGGCAAGGCAGGCTCCTGGCAAACGGAAAGTGCAATTGTCTGGCAAGAGATGCGGA 549

Query:     121 CCACACAGCTGGGACCAGGGCGCTTCCAAATGACCCAGGAAGTGGTTGTGACGAGTGCC 180
             CCAC CAGCTGGG CC GGGCGCTTCCAAATGACCCAGGA GTGGT TG GACGA TGCC
Sbjct:     550 CCACCCAGCTGGGCGCTTGGCGCTTCCAAATGACCCAGGAGGTGGTCTGCGACGAATGCC 609

Query:     181 CTAATGTCAAAGTGTGAATGAAGAACGAACACTAGAAGTGGAAATAGAGCCTGGGGTGA 240
             CTAATGTCAAAGTGTGAATGAAGAACGAAC CT GAAGT GAAATAGAGCCTGGGGTGA
Sbjct:     610 CTAATGTCAAAGTGTGAATGAAGAACGAACGCTGGAAGTAGAAATAGAGCCTGGGGTGA 669

Query:     241 GAGATGGCATGGAGTACCCCTTTATTGGAGAAGGTGAGCCTCATGTGGATGGGGAACCCG 300
             GAGA GGCATGGAGTACCCCTTTATTGGAGAAGGTGAGCCTCA GTGGATGGGGA CC G
Sbjct:     670 GAGACGGCATGGAGTACCCCTTTATTGGAGAAGGTGAGCCTCACGTGGATGGGGAGCCTG 729

Query:     301 GAGACTTACGGTTCCGAATCAAAGTTGTCAAGCACCGGATATTTGAGAGGAGAGGGGATG 360
             GAGA TTACGGTTCCGAATCAAAGTTGTCAAGCAC ATATTTGA AGGAGAGG GATG
Sbjct:     730 GAGATTTACGGTTCCGAATCAAAGTTGTCAAGCACCAATATTTGAAAGGAGAGGAGATG 789

Query:     361 ACCTGTACA 369
             A TGTACA
Sbjct:     790 ATTTGTACA 798

```

Blast-X Results:

>ptnr:SPTREMBL-ACC:Q9UBS4 ERJ3 PROTEIN PRECURSOR - Homo sapiens (Human), 358 aa.

Top Previous Match Next Match
Length = 358

Plus Strand HSPs:

Score = 637 (224.2 bits), Expect = 2.1e-61, P = 2.1e-61
Identities = 119/121 (98%), Positives = 120/121 (99%), Frame = +3

5 Query: 6 KVTLEEVYAGNFVEVVRNKPVARQAPGKRKCNCRQEMRTTQLGPGRFQMTQEVVVCDECPN 185
+VTLEEVYAGNFVEVVRNKPVARQAPGKRKCNCRQEMRTTQLGPGRFQMTQEVVVCDECPN
Sbjct: 139 EVTLEEVYAGNFVEVVRNKPVARQAPGKRKCNCRQEMRTTQLGPGRFQMTQEVVVCDECPN 198

10 Query: 186 VKLVNEERTLEVEIEPGVRDGMIEYFFIGEGEPHVDGEPDGLRFRIKVVKHIFERRGDDL 365
VKLVNEERTLEVEIEPGVRDGMIEYFFIGEGEPHVDGEPDGLRFRIKVVKH IFERRGDDL
Sbjct: 199 VKLVNEERTLEVEIEPGVRDGMIEYFFIGEGEPHVDGEPDGLRFRIKVVKHIFERRGDDL 258

Query: 366 Y 368
Y
Sbjct: 259 Y 259

RISKMARKER7

RISKMARKER7 is a 594 bp novel gene fragment, which has 65% sequence identity to *Mus musculus* hexokinase II [AJ238540], probable 3' UTR. The nucleic acid sequence was initially identified in a cloned fragment having the following sequence:

1 GGGCCCCACTAAACATACACAAAAGAATAAAAAATGTTTCATTTTAACTTAACTGCTTCCTGGTTTTACAAGGCATAAA
81 TATATAGCATCTCCAACAGCTACCTGTAGATTCTGTTAGTGCAAAACCTTAGAAACCCTCCTGGAGCTCAAAGGCATCCG
161 GACTAGT (SEQ ID NO: 7)

The cloned sequence was assembled into a contig resulting in the following consensus sequence:

1 TTTTTTTTTTTTTTTTTTAAAAAGATTATAAAATGAATTTATTGAGTTTCACACAAGATGCACTTATAAAATTAGTACT
81 GAATGCCATTATGACAGAAAGTGAGCATCATCCACTCCCAAGAGCATCTGCAAAGGAAATCAATCTTCAGAGAATAGCA
161 CAGAAACAGAAAATCCAAGCGAACAAAAGATACATCTAGGCCGTGTTCTTGTCTGACCAGGGCCGCATTGGCAAAGC
241 TTTCTCTGCACCTCCCCTGGTTGCCAAGGATACTTTCTTTTGTAAAAAAGTTAGAAAGTGGGGCCCCACTAAAAC
321 ATACACAAAAGAATAAAAAATGTTTCATTTTAACTTAACTGCTTCCTGGTTTTACAAGGCATAAATATATAGCATCTCCA
401 ACAGCTACCTGTAGATTCTGTTAGTGCAAAACCTTAGAAACCCTCCTGGAGCTCAAAGGCATCCGGACTAGTTTTGTACT
481 TAAACAGGATACGGGTAAACCACTTAAAATTTGCCATCTCTGCCCAAAGTGTTCATGAGAACTGAGTTTCAGAAAGACA
561 GCATAGGAAAGAGTCAGAAACGGTCACTTTTTT (SEQ ID NO: 8)

Blast-N Results:

>gb:GENBANK-ID:MMU238540|acc:AJ238540 *Mus musculus* mRNA for hexokinase II - *Mus musculus*, 5474 bp.

Top Previous Match Next Match
Length = 5474

Minus Strand HSPs:

Score = 251 (37.7 bits), Expect = 0.045, P = 0.044
Identities = 121/184 (65%), Positives = 121/184 (65%), Strand = Minus / Plus

Query: 184 GTTCGCTTGGATTTT-CTG-TTTCTGTGCTATTCTCTGAAGATT-GATTTCCTTTGCAGA 128
G TC CT G T T CTG TTT TGTG T TTC TGAA TT GA T C T T CA A
Sbjct: 5287 GCTCTCTCTGCTAATGCTGCTTTGTGTGATCTTCAGTGAACCTTTGACT-CATCT-CATA 5344

5 Query: 127 TGCTCTTGGGAGTGTGGATGATGCTCACTTCTGTGTCATAATGG-CATTC-AGTACTAATTT 70
T C CT GG A T G T TG C TT TGTCAT ATG CA T AG ACTA TT
Sbjct: 5345 TCC-CTGGGCACTCGGTCTAGTGAGCGTTT-TGTCATCATGTACAGTAGAGAACTAGTTG 5402

10 Query: 69 TATAAGTGCATCTTGTGTGAAACTCA-ATAAATTCATTTTATAATCTTTTTTAAAAAAA 11
AT A CAT T TGT AA CT AT AAT AATTTTA T TTTTTT AAAAAA
Sbjct: 5403 AATTAAC-CATGTGATGTTAA-CTATTATTAATA-AATTTTAACCTTTTTTTTCAAAAAA 5459

Query: 10 AAAAAAAAAA 1
AAAAAAAAAA
15 Sbjct: 5460 AAAAAAAAAA 5469

Score = 250 (37.5 bits), Expect = 0.051, P = 0.049
Identities = 122/184 (66%), Positives = 122/184 (66%), Strand = Minus / Plus

20 Query: 184 GTTCGCTTGGATTTT-CTG-TTTCTGTGCTATTCTCTGAAGATT-GATTTCCTTTGCAGA 128
G TC CT G T T CTG TTT TGTG T TTC TGAA TT GA T C T T CA A
Sbjct: 5287 GCTCTCTCTGCTAATGCTGCTTTGTGTGATCTTCAGTGAACCTTTGACT-CATCT-CATA 5344

25 Query: 127 TGCTCTTGGGAGTGTGGATGATGCTCACTTCTGTGTCATAATGG-CATTC-AGTACTAATTT 70
T C CT GG A T G T TG C TT TGTCAT ATG CA T AG ACTA TT
Sbjct: 5345 TCC-CTGGGCACTCGGTCTAGTGAGCGTTT-TGTCATCATGTACAGTAGAGAACTAGTTG 5402

30 Query: 69 TATAAGTGCATCTTGTGTGAAACTCA-ATAAATTCATTTTATAATCTTTTTT-AAAAAA 12
AT A CAT T TGT AA CT AT AAT AATTTTA T TTTTTT AAAAAA
Sbjct: 5403 AATTAAC-CATGTGATGTTAA-CTATTATTAATA-AATTTTAACCTTTTTTTTCAAAAAA 5459

Query: 11 AAAAAAAAAA 1
AAAAAAAAAA
35 Sbjct: 5460 AAAAAAAAAA 5470

Blast-X Results:

>ptnr:SPTREMBL-ACC:Q9VIA2 MST84DB PROTEIN - *Drosophila melanogaster*
(Fruit fly), 70 aa.

[Top](#) [Previous Match](#) [Next Match](#)
Length = 70

Plus Strand HSPs:

Score = 66 (23.2 bits), Expect = 2.2, P = 0.88
Identities = 15/48 (31%), Positives = 25/48 (52%), Frame = +3

45 Query: 66 YKISTECHYDRSEHHPHSQEHLQRKS-----IFRE*HRNRKSKRTRK 191
YK+ ++ H R +H P S++ RK I ++ RNRK R ++
Sbjct: 3 YKVHSHVKARMDHSPRSKDRKDRKGRKAHSHKIHKDYSRNRKDHVRVK 50

50 RISKMARKER8

RISKMARKER8 is a 797 bp novel gene fragment, which has 94% amino acid identity (79% nucleic acid sequence identity) to human GT335 mRNA (ES1 Protein Homolog) [U53003]. The nucleic acid was initially identified in a cloned fragment having the following sequence:

1 CCTAGGACTGCACAACGTGAGTCCTTGAACCAGGCTCTGGAAAAGGTGCCAGACCACCCAATGGGGACACACAGTGAGG

81 CCAGCCCCCAGTGAAATTCCTGCTGCTACCTGGGGCCCTTGGTGAGACTCTGGCTTCCGGCTGGTAGAAGCCAAGGTGG

161 ACGCATAGTTGCAAAGCTCCTCCTTCAGGCACAAAGTGTCTATGCTTCTAATAGAACAGCAGCTCCCGTGTCTGGCTGA
241 CCGGAGCACACAGGCTGAGCGTGCCACAGCGACGACGGAGGCCAAGCGTGGTGTGGTGTACTTTCCCGTGAGTTC
321 CAGCACCTTCTTCACCATGG (SEQ ID NO: 9)

The cloned sequence was assembled into a contig resulting in the following consensus sequence:

1 TTTTTTTTTTTTTTTTTTTTTTTGAGTTTCCACTGTGGAAAAGAGTTTATTGTATGGCTGCAGGGATCTACTACAGAATCC
81 CCCTGGCTGCAGTTAGCTGTGCTTACTCTGGACATATCTCCGAAGACTTGGAGCCTAAATGGTTTTCTTCTTTAGAGCT
161 TTAGTACCCGATCCATCAGACCTAGGACTGCACAACGTGAGTCCTTGAACCAGGCTCTGGAAGAGTGCCAGACCACCC
241 AATGGGGACACACAGTGAGGCCAGCCCCAGTGAAATCCTGCTGTACCTGGGGCCCTTGGTGAGACTCTGGCTTCCGG
321 CTGGTAGAAGCCAAGGTTGGACGCATAGTTGCAAAGCTCCTCCTTCAGGCACAAAGTGTCTATGCTTCTAATAGAACAGC
401 AGCTCCCGTGTCTGGCTGACCGAGCACACAGGCTGAGCGTGCCACAGCGACGACGAGGCCAAGCGTGGTGTGGTGG
481 TGTACTTTCCCGTGAGTTCCAGCACCTTCTTCACCATGGCCCCAATCCCGTCGTGGATGTGGTGGAGTTCCGGTCTCACA
561 CATGAAGGCCGGGGTGGTGACCACCTTGTTTTCTGGTCGACGTGAGCTTCGGTCACACCCCTTCACACAGTGCTTGGCAC
641 CCAGGGCTTTGACGGCTTCCGCGGTTCCAGCATATGGCCACTTGCCCCCTCCTCTTGCTCATGGCCCAGGTGACCTCC
ACACCTTTGATCACTTGGCTGCGAGGACAGGAGCGATGCAGCATAGGCCAATGGGCTTCTTGGCTCCGTGGAATTC
721 (SEQ ID NO: 10)

Blast-N Results:

>gb:GENBANK-ID:HSU53003|acc:U53003 Human GT335 mRNA, complete cds - Homo sapiens, 1652 bp.

Top Previous Match Next Match
Length = 1652

Minus Strand HSPs:

Score = 1141 (171.2 bits), Expect = 7.9e-46, P = 7.9e-46
Identities = 307/385 (79%), Positives = 307/385 (79%), Strand = Minus / Plus

Query: 797 GAATTCCACGAGCCAAGAAGCCCATTTGGCCTATGCTGCATCGCTCCTGTCTCGCAGCC 738
GA TTCCAC GCC GAAGCCCAT GGC T TGCTGCAT GC CCTGTCTCGC GCC

Sbjct: 577 GAGTTCCACCAGGCCGGGAAGCCCATCGGCTTGTGCTGCATTGCACCTGTCTCGCGGCC 636

Query: 737 AAAGTGATCAAAGGTGTGGAGGTACCCGTGGGCCATGAGCAAGAGGAGGGGGCAAGTGG 678
AA GTG TCA AGG GT GAGGT AC GTGGGCCA GAGCA GAGGA GG GGCAAGTGG

Sbjct: 637 AAGGTGCTCAGAGGCGTCGAGGTGACTGTGGGCCACGAGCAGGAGGAAGGTGGCAAGTGG 696

Query: 677 CCATATGCTGGAACCGCGGAAGCCGTCAAAGCCCTGGGTGCCAAGCACTGTGTGAAGGGT 618
CC TATGC GG ACCGC GA GCC TCAA GCCCTGGGTGCCAAGCACTG GTGAAGG

Sbjct: 697 CTTATGCGCGGACCGCAGAGGCCATCAAGGCCCTGGGTGCCAAGCACTGCTGAAGGAA 756

Query: 617 GTGACCGAAGCTCACGTGACCCAGAAAAACAAGTGGTCAACCCCCGCGCTTCATGTGT 558
GTG CGAAGCTCACGT GACCAGAAAAACAAGTGGTCAAC ACCCC GCCTTCATGTG

Sbjct: 757 GTGGTCGAAGCTCACGTGGACCCAGAAAAACAAGTGGTCAACCCCCAGCCTTCATGTGC 816

Query: 557 GAGACCGAAGTCCACCATCCACGACGGGATTGGGGCCATGGTGAAGAAGGTGCTGGAA 498
GAGAC G ACTCCAC ACATCCA GA GGGAT GG GCCATGGTGA GAAGGTGCTGGAA

Sbjct: 817 GAGACGGCACTCCACTACATCCATGATGGGATCGGAGCCATGGTGAGGAAGGTGCTGGAA 876

Query: 497 CTCACGGGAAAGTAACAC-CACC-A-GCACCAC-GCTTGGCCTCCGT-CGTCGCTGTGGC 443
 CTCAC GGAAAGT AC C CA A G C C GCT GGC C G C T GC T C
 Sbjct: 877 CTCACTGGAAAGTGACGCGCATGGACGGGGCCAGCTAGGCGCCAGGACTTGGCC-T--C 933

5 Query: 442 ACGCTCAGCCTGTGT-GCTC-CGGTCAGC 416
 AC CTC G CTG G GCT CGG C GC
 Sbjct: 934 ACCCTCTGGCTGAGGAGCTGTCGG-CTGC 961

Blast-X Results:

>ptnr:SWISSNEW-ACC:P30042 ES1 PROTEIN HOMOLOG, MITOCHONDRIAL PRECURSOR
 (PROTEIN KNP-I) (GT335 PROTEIN) - Homo sapiens (Human), 268 aa.

Top Previous Match Next Match
 Length = 268

Minus Strand HSPs:

Score = 505 (177.8 bits), Expect = 2.0e-47, P = 2.0e-47
 Identities = 94/104 (90%), Positives = 99/104 (95%), Frame = -1

Query: 797 EFHGAKKPIGLCCIAPVLAALKVIKGVEVTVGHEQEEGGKWPYAGTAEAVKALGAKHCVKG 618
 EFH A KPIGLCCIAPVLAALKV++GVEVTVGHEQEEGGKWPYAGTAEA+KALGAKHCVK
 Sbjct: 165 EFHQAGKPIGLCCIAPVLAALKVLRGVEVTVGHEQEEGGKWPYAGTAEAIKALGAKHCVKE 224

Query: 617 VTEAHVDQKNKVVTTPAFMCETELHHIHDGIGAMVKKVLELTGK 486
 V EAHVDQKNKVVTTPAFMCET LH+IHDGIGAMV+KVLELTGK
 Sbjct: 225 VVEAHVDQKNKVVTTPAFMCETALHYIHDGIGAMVRKVLELTGK 268

Principle components analysis was used to generate three eigenvectors used to transform the original expression level data matrix, as shown in Table 4 below. Eigenvector 1 values represent NSAIDs with low risk of hepatotoxicity, Eigenvector 2 values represent NSAIDs with very low risk of hepatotoxicity, and Eigenvector 3 values represent NSAIDs with overdose risk of hepatotoxicity.

Table 4: Transform Eigenvectors for Hepatotoxicity Markers by Risk Classification

Gene	Eigenvector 1	Eigenvector 2	Eigenvector3
RISKMARKER1	26.9	6.7	-0.9
RISKMARKER2	23.3	-1.4	1.5
RISKMARKER3	-26.0	-1.5	-2.3
RISKMARKER4	12.6	-2.2	-6.4
RISKMARKER5	18.0	-1.3	-3.1
RISKMARKER6	-13.8	4.71	19.3
RISKMARKER7	-29.7	-7.5	1.3
RISKMARKER8	19.3	1.2	-2.6
% of variation explained	99.6	0.4	0.1

These eigenvectors may be used to transform the expression levels of RISKMARKERS 1-8 ("RISKMARKERS") in response to a given drug, in order to determine that drug's hepatotoxicity risk. For example, expression levels of RISKMARKERS correlating with

Eigenvector 1 indicates that the test drug has a low risk of hepatotoxicity. Alternatively, a drug's RISKMARKERS expression profile can be generated simultaneously with the above-described training set (or an equivalent set) run in parallel with the test drug, and expression levels associated with the test drug directly compared to those of the training set.

5 A second training set based on type of injury (hepatocellular damage, cholestasis, elevated transaminase level) was also constructed, utilizing the compounds indicated in Table 5, below.

Table 5: Training Set of NSAIDs by Injury Type

Control	Hepatocellular	Cholestasis	Elevated transmainases
Sterile water	Acetaminophen	Benoxaprofen	Zomepirac
10% ethanol	Flurbiprofen	Nabumetone	Mefenamic acid
Canola oil	Ketoprofen	Sulindac	Tenoxicam

10 This analysis produced ten fragments that significantly ($p=8.7 \times 10^{-18}$) discriminated among the drugs in the test set. The identities of these ten fragments (INJURYMARKER 1-10) that are included in the discriminatory set (with GenBank accession numbers) are shown below. Where appropriate, the cloned sequence from isolation is provided, and this sequence was then extended using either Genbank rat ESTs or from internally sequenced (Curagen Corporation) rat fragments. The fragments were used to extend the cloned sequence, and the extended contig
15 sequence is provided as "consensus." Finally, the best BlastN and BlastX results are also provided. In some instances the cloned sequence is identical to a known rat gene, in those instances the name of the gene and the GenBank accession number is provided.

INJURYMARKER1

20 INJURYMARKER1 is a 1025 bp rat express sequence tag (EST) [AI169175]. The nucleic acid was initially identified in a cloned fragment having the following sequence:

```

1   CTGATTTCAAATTTTATTAGAGAACACTTTCGGATTTCAAATTTTATTACAGAACAAACATTTTCTGATTTCAAATTT
81  CTATTATAATTCTCCAGTAATCAAAGCAGTGGCGTTGGCATGAAGGCAGACAGAGGTCATGGAAGAGACCAGGCTCAGAA
161 ACAGCCCCACCATGCACAGCGGGATGTTTTCCCAACAGGGCAACATGCAAAGCCAGGTATCCACATGGGTAGAGTAGAA
241 AGTCAGACCTTACATCTCACACACAAATGAACTCAAATATACCAGAGAGCAAAGCTAAGAGCTAAATCAAGTTTCCTA

```

```
321 GGGCAAGCTGTAGTAGGCTCCCTTGGGTGGGTAAATGCTTTTGTGGATGTGACTACCAAAAATTCAACCAGAGCCAACGA
401 CCCAACTATTAATGGGCAGTGGACCTAAAGAGATTTCTTCAAACGATATATAAAGAAGGCCACCAAGCATATAAAACATG
481 TGACATCAGTAGTCAGAGAGATGGGAAGCAGAAGCACTAGCAGATCTTAACACCTACTAGAACANCCACTAAAAAGAGT
561 AAGACTCACAAGGACATGGGCACTTCTAATCTCTGTGCACTGCTGCCAGGACATACAATAGTGTGGTCACATGGAGACT
641 ACGGCAGTGCCTACTAATAACAGCAGAGTTACCCTAAGACATACAATCTGCTGCGTGTATGCTAAGCAGGATCCGAGGGA
TATTTGTATATACATGTTACAGCATAGTCAGGAGCTCCAGGGTGGGAACAACCTGAGGTACC
721 (SEQ ID NO: 11)
```

The cloned sequence was assembled into a contig resulting in the following consensus sequence:

```
1 CTGATTCAAATTTTATTATAGAACACTTTCTGATTCAAATTTTATTACAGAACAAACATTTTCTGATTCAAATTT
81 CTATTATAATTCTCCAGTAATCAAAGCAGTGGCGTTGGCATGAAGGCAGACAGAGGTCATGGAAGAGACCAGGCTCAGAA
161 ACAGCCCCACCATGCACAGCGGGATGTTTTCCCAACAGGGCAACATGCAAAGCCAGGTATCCACATGGGTAGAGTAGAA
241 AGTCAGACCTTACATCTCACACACAAATGAACTCAAATATACAGAGAGCAAAGCTAAGAGCTAAAATCAAGTTTCCTA
321 GGGCAAGCTGTAGTAGGCTCCCTTGGGTGGGTAAATGCTTTTGTGGATGTGACTACCAAAAATTCAACCAGAGCCAACGA
401 CCCAACTATTAATGGGCAGTGGACCTAAAGAGATTTCTTCAAACGATATATAAAGAAGGCCACCAAGCATATAAAACATG
481 TGACATCAGTAGTCAGAGAGATGGGAAGCAGAAGCACTAGCAGATCTTAACACCTACTAGAACAGCCACTAAAAAGAGT
561 AAGACTCACAAGGACATGGGCACTTCTAATCTCTGTGCACTGCTGCCAGGACATACAATAGTGTGGTCACATGGAGACT
641 ACGGCAGTGCCTACTAATAACAGCAGAGTTACCCTAAGACATACAATCTGCTGCGTGTATGCTAAGCAGGATCCGAGGGA
721 TATTTGTATATACATGTTACAGCATAGTCAGGAGCTCCAGGGTGGGAACAACCTGAGGTACCCACGGCTGGATGAGTAGG
801 TAACAAGAAACATACAGCATACATACAACACACACTAAAGTCTAAAGTACTATTTGTCTTACAAAGGAACTCATACAT
881 GATACAAGCCTTCACGGCATTCTGTACATGAACACGCACACACACACACACACACACACACACACGCCTGAGAATC
TATGTATACCAGGCACTTAGGGTACTCAAATTCAGAAACAGGACAGAGAATGGTGATTGCCATGG
961 (SEQ ID NO: 12)
```

5 Blast-N Results:

>gb:GENBANK-ID:AI169175|acc:AI169175 EST215009 Normalized rat kidney, Bento Soares Rattus sp. CDNA clone RKIBO44 3' end, mRNA sequence - Rattus sp., 670 bp (RNA).

Top Previous Match Next Match
Length = 670

Plus Strand HSPs:

Score = 3305 (495.9 bits), Expect = 4.3e-143, P = 4.3e-143
Identities = 661/661 (100%), Positives = 661/661 (100%), Strand = Plus / Plus

```
Query:      4 ATTTCAAATTTTATTATAGAACACTTTCTGATTCAAATTTTATTACAGAACAAACAT 63
            |||
Sbjct:      1 ATTTCAAATTTTATTATAGAACACTTTCTGATTCAAATTTTATTACAGAACAAACAT 60

Query:      64 TTTCTGATTTCAAATTTCTATTATAATTCACAGTAATCAAAGCAGTGGCGTTGGCATGA 123
            |||
```

Sbjct: 61 TTTCTGATTTCAAATTTCTATTATAATTCTCCAGTAATCAAAGCAGTGGCGTTGGCATGA 120

Query: 124 AGGCAGACAGAGGTCATGGAAGAGACCAGGCTCAGAAACAGCCCCACCATGCACAGCGGG 183

5 Sbjct: 121 AGGCAGACAGAGGTCATGGAAGAGACCAGGCTCAGAAACAGCCCCACCATGCACAGCGGG 180

Query: 184 ATGTTTTCCCAAGGGCAACATGCAAAGCCAGGTATCCACATGGGTAGAGTAGAAAGT 243

10 Sbjct: 181 ATGTTTTCCCAAGGGCAACATGCAAAGCCAGGTATCCACATGGGTAGAGTAGAAAGT 240

Query: 244 CAGACCTTACATCTCACACACAAATGAACTCAAATATACCAGAGAGCAAAGCTAAGAGC 303

Sbjct: 241 CAGACCTTACATCTCACACACAAATGAACTCAAATATACCAGAGAGCAAAGCTAAGAGC 300

15 Query: 304 TAAATCAAGTTTCCTAGGGCAAGCTGTAGTAGGCTCCCTTGGGTGGGTTAATGCTTTTG 363

Sbjct: 301 TAAATCAAGTTTCCTAGGGCAAGCTGTAGTAGGCTCCCTTGGGTGGGTTAATGCTTTTG 360

20 Query: 364 TGGATGTGACTACCAAAAATTCAACCAGAGCCAACGACCCAACCTATTAAATGGGCAGTGGA 423

Sbjct: 361 TGGATGTGACTACCAAAAATTCAACCAGAGCCAACGACCCAACCTATTAAATGGGCAGTGGA 420

Query: 424 CCTAAAGAGATTTCTTCAAACGATATATAAAGAAGGCCACCAAGCATATAAACATGTGA 483

25 Sbjct: 421 CCTAAAGAGATTTCTTCAAACGATATATAAAGAAGGCCACCAAGCATATAAACATGTGA 480

Query: 484 CATCAGTAGTCAGAGAGATGGGAAGCAGAAGCACTAGCAGATCTTAACACCTACTAGAAC 543

30 Sbjct: 481 CATCAGTAGTCAGAGAGATGGGAAGCAGAAGCACTAGCAGATCTTAACACCTACTAGAAC 540

Query: 544 AGCCACTAAAAAGAGTAAGACTCACAAGGACATGGGCACCTTCTAATCTCTGTGCACTGC 603

Sbjct: 541 AGCCACTAAAAAGAGTAAGACTCACAAGGACATGGGCACCTTCTAATCTCTGTGCACTGC 600

35 Query: 604 TGCCAGGACATACAATAGTGTGGTCACTATGGAGACTACGGCAGTGCCTACTAATAACAG 663

Sbjct: 601 TGCCAGGACATACAATAGTGTGGTCACTATGGAGACTACGGCAGTGCCTACTAATAACAG 660

40 Query: 664 C 664

Sbjct: 661 C 661

INJURYMARKER2

INJURYMARKER2 is a 893 nucleotide sequence encoding phosphotidylethanolamine N-methyltransferase [L14441]:

45 1 tccccgctga gttcatcacc agggacaggt gacctgagct gccctggag ccagctccc

61 atttccttct ggttctggcc gatctcttcg ttatgagctg gctgctgggt tacgtggacc

121 ccacagagcc cagctttgtg gcggctgtgc tcaccattgt gttcaatcca ctcttctgga

181 atgtggtagc aaggtgggag cagagaactc gcaagctgag cagagccttc gggccccctt

241 acctagcctg ctattccctg ggcagcatca tcctgcttct gaacatcctc cgctcccact

50 301 gcttcacaca ggccatgatg agccagccca agatggaggg cctggatagc cacaccatct

361 acttcctggg ccttgcactc ctgggctggg gactcgtgtt tgtgctctcc agcttctatg

421 cactgggggt cactgggacc tttctaggtg actacttttg gatcctcaag gagtccagag

481 tgaccacatt tcccttcagc gtgctggaca accccatgta ctggggaagt acagccaact

541 acctaggctg ggcacttatg cagccagacc ctacaggcct gctgttgacg gtgctggtgg

55 601 cactcgtcta cgtggttgc ctctgtttg aagagccctt cactgcggag atctaccggc

661 ggaaagccac caggttgac aaaaggagct gacagggcca tgagggacct ttggaaagcc

721 ggattgctc ccggctgacc caagcaacaa cccttctcgg ggagagcagc gctggccatt

781 gtacctgtgc cttggaaacc agtcatgggg gtgctcaggc attatgtcat gtgactgctg

841 agaccccat cccaccaat ccctgacaca ctaataaagg ctttgtgacc tcc

INJURYMARKER3

INJURYMARKER3 is a 1131 nucleotide hexokinase-encoding sequence [M86235]:

5	1	agcaggaatc	ccctccgctt	gcgggtagga	agcttgggga	gcagcctcat	ggaagagaag
	61	cagatcctgt	gcgtggggct	ggtggtgctg	gacatcatca	atgtggtgga	caaataccca
	121	gaggaagaca	cggatcgcag	gtgcctatcc	cagagatggc	agcgtggagg	caacgcgtcc
	181	aactcctgca	ctgtgctttc	cttgctcgga	gcccgtgtg	ccttcatggg	ctcgttggcc
	241	catggccatg	ttgccgactt	cctggtggcc	gacttcaggc	ggaggggtgt	ggatgtgtct
10	301	caagtggcct	ggcagagcca	gggagatacc	ccttgctcct	gctgcatcgt	caacaactcc
	361	aatggtcccc	gtaccattat	tctctacgac	acgaacctgc	cagatgtgtc	tgctaaggac
	421	tttgagaagg	tcatctgac	ccggttcaag	tggatccaca	ttgagggccg	gaatgcacgc
	481	gaacaggtaa	agatgctaca	gcggatagaa	cagtacaatg	ccacgcagcc	tctgcagcag
	541	aagggtccggg	tgtccgtgga	gatagagaag	ccccgagagg	aactcttcca	gctgttccgc
15	601	tatggagagg	tgggtgttgt	cagcaaagat	gtggccaagc	acctgggggt	ccggtcagca
	661	ggggaggccc	tgaagggtct	gtacagtcgt	gtgaagaaag	gggtacgct	catctgtgcc
	721	tgggctgagg	agggagccga	tgccctgggc	cccgcggcc	agctgctcca	ctcagatgcc
	781	ttcccaccac	cccagtagt	agacactctc	ggggctggag	acaccttcaa	tgctctgtgc
	841	atcttcagcc	tctccaaggg	aaacagcatg	caggaggccc	tgagattcgg	gtgccagggtg
20	901	gctggcaaga	agtgtggctt	gcaggggttt	gatggcattg	tgtgagagat	gagcgggtggg
	961	aggtagcagc	tgcacacctc	agaggctggc	accactgcct	gccattgcct	tcttcatttc
	1021	atccagcctg	gcgtctggct	gccagttcc	ctgggccagt	gtaggctgtg	gaacgggtct
	1081	ttctgtctct	tctctgcaga	cacctggagc	aaataaatct	ttccctgagc	c

INJURYMARKER4

INJURYMARKER4 is a 1994 nucleotide sequence encoding mitochondrial HMG-CoA Synthase [M33648]:

	1	atctctccca	ggggctgtgg	actgctggct	ttctgttgat	accttagaga	tgcagcggct
	61	tttggctcca	gcaaggcggg	tctgcaagt	gaagagagtc	atgcaggaat	cttcgctctc
30	121	accgcctcac	ctgctccccg	cagcccagca	gagggtttct	acaatccctc	ctgctccctc
	181	ggccaaaact	gatacatggc	caaaagatgt	gggcacctct	gccttgagg	tctactttcc
	241	agcccaatat	gtggacaaa	ctgacctgga	gaagttcaac	aatgtggaag	cagggaagta
	301	cacagtgggc	ttgggcccga	ccgctatggg	cttctgttcg	gtccaggagg	acatcaactc
	361	cttgtgcctc	acagtgggtc	agaggctgat	ggaacgcaca	aagctgcat	gggatgccgt
35	421	aggccgcctg	gaagtgggca	cggaaacat	cattgacaag	tccaaggctg	tcaagacagt
	481	gctcatggag	ctcttccagg	attcaggcaa	cactgacatc	gagggcatag	ataccaccaa
	541	cgctgctat	ggtggcactg	cctccctctt	caacgctgcc	aactggatgg	agtccagcta
	601	ctgggatggg	cgctatgccc	tgggtggtctg	tggatgatct	gcagtctacc	caagtggtaa
	661	ccccgcctcc	acaggtgggtg	ccggggctgt	ggcaatgctg	attgggcccc	aggccccgct
40	721	agtcctggaa	caagggtgta	ggggaaccca	catggagaac	gcctatgact	tctacaaacc
	781	aaacttggcc	tcagagtatc	cactggtgga	tgggaagctg	tctatccagt	gctacctgct
	841	ggccttggac	cgatgctatg	cagcttaccg	caggaaaatc	cagaatcagt	ggaagcaagc
	901	tggaaacaac	cagcctttca	ccctcgatga	cgtgcaatat	atgatcttcc	acacaccctt
	961	ttgcaagatg	gtccagaaat	ccctagctcg	gctgatgttc	aatgacttcc	tgtcatctag
45	1021	cagtgacaag	cagaacaact	tatacaaggg	tctagaggcc	ttcaagggtc	taaagctgga
	1081	agaaaacctac	accaacaagg	atgttgacaa	ggctctgctg	aaggcctccc	tggacatgtt
	1141	caacaagaaa	accaaggcct	ccctttacct	ctccacaaac	aatgggaaca	tgtacacctc
	1201	gtccctctac	gggtgcctgg	cctcacttct	ctcccaccac	tctgcccagg	aattggccgg
	1261	ctccaggatt	ggagccttct	cctacggctc	aggcttagca	gcaagtttct	tctcatttgc

	1321	agtgtccaag	gacgcttccc	cagggtcccc	tctggagaag	ctggtgtcta	gtgtgtcaga
	1381	tctgccccaa	cgtctagact	cccggagacg	catgtccctt	gaggaattca	cagaaataat
	1441	gaatcagaga	gagcaatttt	accacaaggt	gaactttctt	ccccctggtg	acacaagcaa
	1501	cctcttccca	ggcacttggg	accttgaacg	agtggatgag	atgcaccgca	gaaaatatgc
5	1561	ccggcgctcc	gtctaaggag	accaatccat	acaaccattc	cccggggaaa	gaatgtgagc
	1621	agagccgtta	cccaaacggc	ttccacttaa	aattccaccc	acagcagtga	acggtgaata
	1681	gacacagcga	ccccatagga	tctgtcccgc	gggtgaagggc	ctccctctgt	ggatcctggg
	1741	tgaccctccc	tgaagcagt	agcaccacag	gttctgtctg	ggaccagagc	ccccctgtgg
	1801	agagggagaa	agaaaagggg	gccgctgacc	tgcagggata	cagaccttcc	ccacagcctg
10	1861	gcagccgccc	gtttgttgca	gcttattatc	agactgtggg	ctatcatagt	tcatgtctgt
	1921	ttcttaaagt	ttcccgagaa	tttctaaaat	tttgtatcta	aacttttaat	atggcgatta
	1981	aaaggagaga	agga				

INJURYMARKER5

15 INJURYMARKER5 is a 1850 nucleotide sequence encoding cathepsin C [D90404],
having the following nucleic acid sequence:

	1	gaattccggg	tctagttggt	gttttctctg	ccatctgctc	tccgggcgcc	gtcaaccatg
	61	ggtcogtgga	cccactcctt	gcgcgcgcgc	ctgctgctgg	tgcttttggg	agtctgcacc
	121	gtgagctcog	acactcctgc	caactgcact	taccctgacc	tgctgggtac	ctgggttttc
20	181	cagggtgggc	ctagacatcc	ccgaagtcac	attaactgct	cggtaatgga	accaacagaa
	241	gaaaaggtag	tgatacacct	gaagaagttg	gatactgcct	atgatgaagt	gggcaattct
	301	gggtattttca	ccctcattta	caaccaaggc	tttgagattg	tggtgaatga	ctacaagtgg
	361	tttgcgTTTT	tcaagtatga	agtcaaaggc	agcagagcca	tcagttactg	ccatgagacc
	421	atgacagggg	gggtccatga	tgtcctgggc	cggaaactggg	cttgctttgt	tggcaagaag
25	481	atggcaaata	actctgagaa	ggtttatgtg	aatgtggcac	accttggagg	tctccaggaa
	541	aaatattctg	aaaggctcta	cagtcacaac	cacaactttg	tgaaggccat	caattctggt
	601	cagaagtctt	ggactgcaac	cacctatgaa	gaatatgaga	aactgagcat	acgagatttg
	661	ataaggagaa	gtggccacag	cggaggatc	ctaaggccca	aacctgcccc	gataactgat
	721	gaaatacagc	aacaaatttt	aagtttgcca	gaatcttggg	actggagaaa	cgtccgtggc
30	781	atcaattttg	ttagccctgt	tcgaaacca	gaatcttgtg	gaagctgcta	ctcatttgcc
	841	tctctgggta	tgctagaagc	aagaattcgt	atattaacca	acaattctca	gaccccaatc
	901	ctgagtcctc	aggaggttgt	atcttgtagc	ccgtatgccc	aaggttggtg	tgggtgattc
	961	ccatacctca	ttgcaggaaa	gtatgcccac	gattttgggg	tgggtggaag	aaactgcttt
	1021	ccctacacag	ccacagatgc	tccatgcaaa	ccaaaggaaa	actgcctccg	ttactattct
35	1081	tctgagtact	actatgtggg	tggtttctat	gggtggctgca	atgaagccct	gatgaagcct
	1141	gagctgggtca	aacacggacc	catggcagtt	gcctttgaag	tccacgatga	cttctctgac
	1201	taccacagtg	ggatctacca	ccacactgga	ctgagcgacc	ctttcaaccc	ctttgagctg
	1261	accaatcatg	ctgttctgct	tgtgggctat	ggaaaagatc	cagtcactgg	gttagactac
	1321	tggattgtca	agaacagctg	gggctctcaa	tgggggtgaga	gtggctactt	ccggatccgc
40	1381	agagggaactg	atgaatgtgc	aattgagagt	atagccatgg	cagccatacc	gattcctaaa
	1441	ttgtaggacc	tagctcccag	tgtcccatac	agctttttat	tattcacagg	gtgatttagt
	1501	cacaggctgg	agactttttac	aaagcaatat	cagaagctta	ccactaggta	cccttaaaga
	1561	attttgcctt	taagtttaaa	acaatccttg	atttttttct	tttaatatcc	tccctatcaa
	1621	tcaccgaact	actttttcttt	ttaaagtact	tgggttaagta	atacttttct	gaggattggg
45	1681	tagatattgt	caaatatttt	tgtctggctac	ctaaaatgca	gccagatgtt	tcattgttaa
	1741	aaatctatat	aaaagtgcac	gctgcctttt	ttaaattaca	taaatcccat	gaatacatgg
	1801	ccaaaatagt	tatttttttaa	agacttttaa	ataaatgatt	aatcgatgct	

INJURYMARKER6

INJURYMARKER6 is a 993 nucleotide sequence encoding hydroxysteroid sulfotransferase [D14989]:

1	ggcaagggct	ggaatactaa	aagtatttca	tgatgtcaga	ctataacttg	tttgaaggaa
5	61	taccttttcc	tgcccttttg	ttttccaaag	aaattctgga	aaatagttgt
	121	tggtaaaaga	agacgacttg	atcatattga	cttaccctca	gtcaggaacg
	181	tcgagattgt	ctgcttgatt	cagaccaagg	gagatcccaa	gtggatccaa
	241	tctgggatcg	ctcaccctgg	atagagactg	gttcaggata	tgataaatta
	301	aaggaccacg	actcatgacc	tcccatcttc	ccatgcatct	tttctccaag
10	361	gttccaaggc	caaggtgata	tatctcatca	gaaatcccag	agatgttctt
	421	atTTTTTctg	gagtaagatc	gccctggaga	agaaaccaga	ctcgctggga
	481	aatgggttct	caaaggaaat	gttgcataatg	gatcatgggt	tgagcacatc
	541	tgtctatgag	agaatgggac	aacttcttgg	tactgtacta	tgaagacatg
	601	caatgggatc	cataaagaag	atatgtgact	tcctggggaa	aaaattagag
15	661	tgaatttggg	cctcaagtat	agttccttcc	aagtctgtga	agaaaacaac
	721	atagcctcat	ggagaaggaa	ctgattctta	ctggttttac	tttcatgaga
	781	ctaagtactg	gaagaatcac	ttcacagtag	ccaagctga	agcctttgat
	841	aggagaaaat	ggcgggtttc	cctccaggga	tgttcccatg	ggaataaatt
	901	ttaaatatatt	tatgaacact	gatgtttatg	tttatgttgt	tctatgatgt
20	961	gaatgtgatc	attgaataaa	tcctgttgtg	gat	

INJURYMARKER7

INJURYMARKER7 is a 5001 nucleotide sequence encoding insulin-like growth factor binding protein [L22979]:

25	1	cacaaaccca	gcgagcattg	aacactgcac	acggccatct	gccagagag	ctgtgaccac
	61	cacttccgct	actatctact	cagaaagtcg	tgactactga	gccactgctg	cctgcccaga
	121	ttctcatcca	ccgcctgctg	cgtctggttg	cgatgccgga	gttcctaact	gttgtttctt
	181	ggcgttccct	gatectcctg	tccttccagg	ttcgcgtagt	cgctggagcc	ccccagccat
	241	ggcactgtgc	tccttgcact	gctgagaggc	tggagctctg	tccaccctg	cctgcttctg
30	301	gcccagagat	ttctcggcct	gcgggctgtg	gctgctgccc	gacatgtgcc	ttgccactgg
	361	gtgctgacct	tgggtgtggc	actgcggcct	gcgctcaggg	actcagctgc	cgtgcgctgc
	421	caggggagcc	tcgacctctg	catgccctca	cccgtggcca	gggagcctgt	gtactagaac
	481	ctgcgcgacc	cgccacgagc	agcttgtccg	gttctcagca	tgaaggtact	acagccctct
	541	ctgcctcttg	atctcttggc	taggacacac	gtgctttcta	ggcacgtcag	aggcctatcc
35	601	ggaacctata	gcagatagga	caaaggctct	ccatgcccac	tttgagcttt	cagcctcaaa
	661	taaggccctc	agttaggctg	tggcggcttg	ggaaacacca	gaggtgtcaa	tccagtagca
	721	gagtggagaa	gttgggaaga	atgttccaag	ctcccagtg	agagtggaga	gttgggaaga
	781	atgttcacag	actaggtagt	actgatcctg	cttgggtctt	cagtggggag	ggagctatgg
	841	ggctgccagg	tgggtgggg	gctggcccaa	acacctctt	ctgtgggtcc	tgaccttggc
40	901	agttccaatg	gctaaaaggt	ccaggaaggt	ttaggatggg	agccctcctg	ctgccccag
	961	gaggtttgca	atgtcctttg	tagcatatat	cctgccacac	agtatgtgct	tcccagatgt
	1021	ttacagaaca	taatgtgaaa	atttaggccc	aaaccttcac	ttccattcat	tgctatagac
	1081	aaacagtgtt	tgaagtgtat	gttgccctg	aggagtctga	caatcaggcg	ctttcctgaa
	1141	tttaagcact	ggtttgtttg	taataggaag	cttgggaaat	gcctcttct	ctgctccagc
45	1201	ccctatctcc	cctgtctggg	ctgcatgcac	ttcctgtgtg	ggttaaggac	ctcatggttc
	1261	catattctga	cgggaagccg	gactgcaggc	atctgatcct	tttgactaaa	tggaaagact

1321	atcccaacggt	tccttagaaa	cggtctccc	caggagcgat	gtctgataat	gtccctctct
1381	gtgaggggct	gcctaagagg	tgtcgggtgt	caagaaagca	gggtcccag	aaaagaagag
1441	gatgggtggt	tgagggtggg	aaggctacac	tctacacctt	gcttctcaac	tatcccccta
1501	ctggggtctt	acgagattct	ttttgtggtg	tggagaggag	agctgagtgg	tcaagtctca
5 1561	ccactaacgg	gttcaagcct	tggcctcagt	ccttggcttc	ttcaggatta	catcctagac
1621	ccaactctct	ctgccatggg	gactcccttg	cctaacccca	aaacatacca	tttcccaga
1681	aaggaattag	tattgctaata	tgggtgataat	tgttcccaaa	tagcccactg	gtgaaaacaa
1741	agcctgattt	cacctgactg	ttacagattg	gtcttaaggc	ggtagacgtg	agtgcacatg
1801	gagtgacacc	tcagggtctca	tcgtctgtgt	ctgtgggggt	cgttttcaga	ggcaaaggct
10 1861	gctgtggcct	ctgaggatga	gcttgccgag	agcccagaga	tgacagagga	acagctgctg
1921	gatagcttcc	acctcatggc	cccatcccg	gaggaccagc	ccatcctgtg	gaatgccatt
1981	agcacctaca	gcagcatgcg	ggcccgggag	atcactgacc	tcaagaaatg	gaaggtgaga
2041	ccctgcactc	agaccttcag	gtttagctat	ctacgtgaag	aggtttgtct	agacgatttc
2101	ttaaagggca	ctgagcatgg	ggctgagaac	gggatataaa	ctaccctcat	ccctgatgta
15 2161	tttctgcctc	cttaaaaaata	tggcaagtat	ctcagagcat	aaggtaggcc	atttttcagt
2221	ctaggtttct	ctgtcaccga	tgcagcagct	tcagtgattg	ttagccacca	accagctcca
2281	cggttttgccc	agccttttagc	tatgcacttt	agctatgcag	ttaaacttctc	tagctttact
2341	ggctgttttt	caacttgacc	acttggggga	gacagagaac	caaagggtgga	gagaaagtac
2401	ggcagaggca	ttgaagaagt	acacctaaag	aaatgaaagg	ataaacattg	ttagggggcac
20 2461	tttagaattt	catatggaaa	ttgtccaaat	cagtgccttg	ttccgtaatc	aatttgacat
2521	acaccaaagt	caaggatggc	tgtttgaaaa	atctaggcat	ttatgatgct	aaattccaca
2581	cacagagact	gagcctgtct	tttttattag	agttcagggt	ctcaagttaa	tcagagatat
2641	ccagggtcag	gaagcattta	taccattggc	caggctctta	ccacaatgtc	gttaaggggg
2701	tctccagaaa	atgccactga	gggaggatga	gagtggtgtc	cctgtccttt	atctacatat
25 2761	cccaagccag	agaccaacct	gtcctgtctca	cagatgggga	aacatctcag	ccgttgtcta
2821	aattgataat	ttttgtctct	tgtactcatg	ctaataataa	attatccttt	taggagccct
2881	gccaacggga	actctataaa	gtgttagaga	gattagctgc	cgctcaacag	aaagcaggag
2941	atgagatcta	caaaatttat	ctgccaaact	gcaacaagaa	tggattttat	cacagcaaac
3001	aggtagggtg	ctttgtctcat	ccagatcctt	gtaaaacttc	atgatttttt	tttttaaagt
30 3061	caaatgattc	acaggcccaa	tacatcatcat	gggtagcttt	cttaggtgag	atccagccct
3121	gcagtagattg	ggagaagcta	gtcctgagaa	agagatagt	tgatggatga	ggaacacttc
3181	agccagaagg	gaggactaag	cattagtgtg	atgagtgagg	agcacttcag	ttaacaggga
3241	ggactaagca	ttagtgtgat	gagtgaggac	cacttcaagc	cagagggagg	actaacattg
3301	gcagtatgat	gagtgaggag	cacttcagcc	agtagggagg	actaaccatt	agtctcatca
35 3361	ctaggagcac	ctcagccaag	tagggaggac	taaccattag	tctcacactc	acccaacatc
3421	ttcagtcagg	actaagcatt	agtgtgatga	gtgaggagca	cttcagtcag	tagggaggac
3481	tacattagtg	tgatgagtga	ggagcacttc	agccagtagg	gaggactaac	cgttcactca
3541	gattagcaga	gatggatgtt	ccatatactg	atgtccaggt	ttcagttcct	cacaactaga
3601	ggaaaaggac	acagtcagtg	taggagacag	atgtctcgcg	ttctctcttc	ccacaaataa
40 3661	aaacaaactc	tgtagtaaga	cacaccaatt	gtgctttgcc	tatcgaataa	tgagattgaa
3721	gaagtcagg	cttaatttcg	acgcaacttt	agcaactcag	gaagtgcagg	ttctggaatt
3781	tcattgagga	aaaacttgag	gtctaggtct	agccgtgtgg	tagagatgg	gagacctatc
3841	gttgagctcc	tttggcagag	ggccatggag	caggtaaccg	tcaaaacaat	ataccactga
3901	gtaaacagat	gagattgtta	tcagggtgtgc	cataaagcca	acctctccgt	tttgtgatga
45 3961	caaccagaag	ggcattggtc	tgccgagcct	tagccagcag	gtagctgtgc	agtgtctggc
4021	ctcactgagg	gacaggggtg	ccagagctct	tacctctctg	tgctcttgac	ctcggtcctg
4081	tctttgcagt	gcgagacatc	tctggatgga	gaagctgggc	tctgctgggt	tgtctaccca
4141	tggagtggga	agaagatccc	tggatctctg	gagaccagag	gggaccccaa	ctgccaccag
4201	tattttaatg	tgcaaaaactg	aaagttgttt	cctccctcct	tcttcacaca	aaatatttaa
50 4261	gtatatagtg	tattttatact	ccggagcaca	ccattttata	tatgtgtata	tgtatatatc
4321	caggaactag	ttttttatact	ccacatgctg	cttgatgtac	aagtgggttt	gtattttatc
4381	actctaagtt	tattttttttc	tacctgtctc	ttgtgtctga	ttaatttata	taactgaagc
4441	ttttctcatc	tccatacatg	taataactac	catctcagct	ctccagagt	ctgtcttga
4501	aagggcagcg	cggtagctgc	ctagaaacag	cacaagtcag	tctgaggtag	gggcctttca
55 4561	gtgggttcag	ggaggaaggt	tagccctggc	tgggggagac	ttcctcatcg	aatcccacag
4621	gtctgtgtct	gatgcctatt	ggctgggaag	gttccgatgt	tgggtgtgta	atcaaagcta
4681	aacgtgga	gctgcgtccc	atgcactgtt			

4741 tggaaacact gctgtctctg tgggaattcca gctctgtgct cattccctca gtccgttcgg
4801 ctttcccgtc cgctgattc ctgggtctgt gctttgggga tagatgttgc aatacagggt
4861 gcttgtttgt ttacagaaca ccctggacaa acactctgtg actttatggc cccattttca
4921 agcagcatca ggcctctgtc tgggccagac tacagagccc ctctccttg gtccatctcc
5 4981 ctttcttccc agggccctca g

INJURYMARKER8

INJURYMARKER8 is 579 bp rat expressed sequence tag (EST) [AA851963]. The nucleic acid was initially identified in a cloned fragment having the following sequence:

1 TGTACATAATTTATTAAAAATGTCTCTGACACAAATAATGACTCCACTGCATACATAGTTGGTGTTCAAAAATTTCCCCA
81 ATGTTTGTCTGGACACAATTGTTATTAGCCAACTCGGTGAATTCAAGACATTGTTCCACACAATGAACAATCGCACACA
161 TGAGAACTGCACCTAGAATGTCCATCCTAGAATCTCCATCCAGTCAAAGTGCTGAGCTCACTGACTGAAGGAAACA
241 TGACCTGTGTTCTAGA (SEQ ID NO: 13)

The cloned sequence was assembled into a contig resulting in the following consensus sequence:

1 TGTACATAATTTATTAAAAATGTCTCTGACACAAATAATGACTCCACTGCATACATAGTTGGTGTTCAAAAATTTCCCCA
81 ATGTTTGTCTGGACACAATTGTTATAAGCCAACTCGGTGAATTCAAGACATTGTTCCACACAATGAACAATCGCACACA
161 TGAGAACTGCACCTAGAATGTCCATCCTAGAATCTCCATCCAGTCAAAGTGCTGAGCTCACTGACTGAAGGAAACA
241 TGACCTGTGTTCTAGAACGTAGCTGGCTATGAAGTTTACTCATGTGTAATTCCTTAAAAAGATTAAATTGTTGGCCCCA
321 TTTCTATATTTATAAAATAACTATAATTACAACTTTCTAAAATAATTTTACAACCATGTAATTATGACTAACCATAT
401 CATCTAAAAAGTAAGTGAAGTCATTGTCTAGAGATTGTCTGAGATTATTCTGCTGAGAAGCTTACTTCAAACCTCTTATC
481 ACTACTTCCTACTTCCAGTGCCTTGAATTAAGAACAGAAATTGTAATCTATGCTATTCTACATCAGATTGACACAACCTA
561 CTTCTAAGTACACTATTGC (SEQ ID NO: 14)

Blast-N Results:

>gb:GENBANK-ID:AA851963|acc:AA851963 EST194732 Normalized rat spleen, Bento Soares
Rattus sp. cDNA clone RSPA086 3' end, mRNA sequence - Rattus sp., 538 bp (RNA).

Top Previous Match Next Match
Length = 538

Plus Strand HSPs:

Score = 2681 (402.3 bits), Expect = 8.1e-115, P = 8.1e-115
Identities = 537/538 (99%), Positives = 537/538 (99%), Strand = Plus / Plus

Query: 42 CTCCACTGCATACATAGTTGGTGTTCAAAAATTTCCCAATGTTTGTCTGGACACAATT 101
 Sbjct: 1 CTCCACTGCATACATAGTTGGTGTTCAAAAATTTCCCAATGTTTGTCTGGACACAATT 60

5 Query: 102 GTTATAAGCCAACTCGGTGAATTCAAGACATTGTTCCACACAATGAACAATCGCACACAT 161
 Sbjct: 61 GTTATTAGCCAACTCGGTGAATTCAAGACATTGTTCCACACAATGAACAATCGCACACAT 120

10 Query: 162 GAGAACTGCACCTAGAATGTCCATCCTAGAATCTCCATCCATCCAGTCAAAGTGCTGAGC 221
 Sbjct: 121 GAGAACTGCACCTAGAATGTCCATCCTAGAATCTCCATCCATCCAGTCAAAGTGCTGAGC 180

15 Query: 222 TCACTGACTGAAGGAAACATGACCTGTGTTCTAGAACGTAGCTGGCTATGAAGTTTACTC 281
 Sbjct: 181 TCACTGACTGAAGGAAACATGACCTGTGTTCTAGAACGTAGCTGGCTATGAAGTTTACTC 240

20 Query: 282 ATGTGTAAATTCTTAAAAAGATTAAATTGTTTGGCCCATTTCTATATTTTCAAAAATAA 341
 Sbjct: 241 ATGTGTAAATTCTTAAAAAGATTAAATTGTTTGGCCCATTTCTATATTTTCAAAAATAA 300

25 Query: 342 CTATAATTACAACTTTCTAAAAATAATTTTACAACCATGTAATTATGACTAACCATATC 401
 Sbjct: 301 CTATAATTACAACTTTCTAAAAATAATTTTACAACCATGTAATTATGACTAACCATATC 360

30 Query: 402 ATCTAAAAAGTAAGTGAAGTCATTGTCTTAGAGATTGTCTGAGATTATTCTGCTGAGAAG 461
 Sbjct: 361 ATCTAAAAAGTAAGTGAAGTCATTGTCTTAGAGATTGTCTGAGATTATTCTGCTGAGAAG 420

35 Query: 462 CTTACTTCAAACCTTATCACTACTTCCACTTCCAGTGTCTTGAATTAAGAACAGAAA 521
 Sbjct: 421 CTTACTTCAAACCTTATCACTACTTCCACTTCCAGTGTCTTGAATTAAGAACAGAAA 480

Query: 522 TTGTAACATGCTATTCTACATCAGATTGACACAACCTACTTCTAAGTACACTATTGC 579
 Sbjct: 481 TTGTAACATGCTATTCTACATCAGATTGACACAACCTACTTCTAAGTACACTATTGC 538

INJURYMARKER9

INJURYMARKER9 is a 2495 nucleotide catalase-encoding sequence[M11670], having the following nucleic acid sequence:

1 attgcctacc cggggtggag accgtgctcg tccggccctc ttgcctcacg ttctgcagct
 61 ctgcagctcc gcaatcctac accatggcgg acagccggga cccagccagc gaccagatga
 121 agcagtggaa ggagcagcgg gccctcaga aaccgatgt cctgaccacc ggaggcggga
 181 acccaatagg agataaactt aatatcatga ctgcggggcc ccgaggggcc ctccctcgttc
 241 aagatgtggt tttcaccgac gagatggcac actttgacag agagcggatt cctgagagag
 301 tggtagatgc aaaggagca ggtgcttttg gatactttga ggtagccac gatattacca
 361 gatactccaa ggcaaagggtg tttgagcata ttgggaagag gactcctatt gccgtccgat
 421 tctccacagt cgctggagag tcaggctcag ctgacacagt tcgtgaccct cgtgggtttg
 481 cagtgaat ctacactgaa gatggtaact gggacctcgt gggaaacaac acccctatctt
 541 tcttcatcag ggaatgcatg ttgtttccat cctttatcca tagccagaag agaaaccac
 601 aaactcacct gaaggaccct gacatggtct gggacttctg gactccttgc ccagagtctc
 661 tccatcaggt tactttcttg ttcagcgacc gagggattcc agatggacat cggcacatga
 721 atggctatgg ctccacacac ttcaagctgg ttaatgcgaa tggagaggca gtgtactgca
 781 agttccatta caagactgac cagggcatca aaaacttgcc tgttgaagag gcaggaagac
 841 ttgcacagga agaccggat tatggcctcc gagatctttt caatgccatc gccagtggca
 901 attacccatc ctggactttt tacatccagg tcatgacttt caaggaggca gaaaccttcc
 961 catttaatcc atttgacctg accaagggtt ggcctcaca ggactaccct cttataaccag
 1021 ttggcaaaact ggtcttaaac agaaatcctg ctaattatct tgctgaagtt gaacagatgg
 1081 cttttgaccc aagcaacatg cccctggca ttgagcccag cccggacaag atgctccagg
 1141 gccgcctttt tgcttaccac gacactcacc gccaccgcct gggaccaaac tatctgcaga

1201 tacctgtgaa ctgtccctac cgtgctcgcg tggccaacta ccagcgcgat ggcctcatgt
1261 gcatgcatga caaccagggg ggtgctccca actactaccc caacagcttc agcgcaccag
1321 agcagcaggg ctgcggccctg gagcaccata gccagtgtct tgcagatgtg aagcgcttca
1381 acagtgtctaa tgaagacaac gtcactcagg tgcggacatt ctatacgaag gtgttgaatg
5 1441 aggaggagag gaaacgcctg tgtgagaaca ttgccaacca cctgaaagat gctcagcttt
1501 tcattcagag gaaagcggtc aagaatttca ctgacgtcca ccctgactac ggggcccagag
1561 tccaggtctt tctggaccag tacaactccc agaagcctaa gaatgcaatt cacacctacg
1621 tacaggccgg ctctcacata gctgccaagg gaaaagctaa cctgtaaagc acgggtgctc
1681 agcctcctca gcctgcactg aggagatccc tcatgaagca gggcacaagc ctcaccagta
10 1741 atcatcgctg gatggagtct cccctgctga agcgcagact cacgctgacg tctttaaaac
1801 gataatccaa gcttctagag tgaatgatag ccatgctttt gatgacattt cccgaggggg
1861 aaattaaaga ttagggctta gcaatcactt aacagaaaca tggatctgct taggacttct
1921 gtttggatta ttcattttaa atgattacaa gaaaggtttt ctagccagaa acatgatttg
1981 attagatatg atatatgata aaatcttggt gattttacta tagtcttatg ttacctcaca
15 2041 gcctggtata tatacaacac acacacacac acacacacac acacacaaa acacacatac
2101 actatacaca cacacacaca cacacactaa aacacacata cacaacacac acatacacta
2161 cacacacaga acacacaaca caaacatata cacataggca cacacacaca cacacacaca
2221 cacacacaca cacacacaca cacacatgaa tgaagggatt ataaagatgg cccaccacaga
2281 attttttttt attttttctaa ggtccttata agaaaaacca tacttggatc atgtcttcca
20 2341 aaaataactt tagcactgtt gaaacttaat gtttattcct gtgtagtga ttggattcct
2401 tttcccttg aaattatgtt tatgctgata cacagtgatt tcacataggg tgatttgtat
2461 ttgcttacat ttttacaata aatgatctt catgg

INJURYMARKER10

INJURYMARKER10 is a 1884 nucleotide betaine homocysteine methyl transferase-encoding sequence [AF038870]:

1 caagcctttg ctggagaccg ctctgttcca gtccgcagct ggcttcagcg ccaactcagga
61 caccggaaaag atggcaccga ttgccggcaa gaaggccaag aggggaatct tagaacgctt
121 aaatgctggc gaagtctgta tgcggagatgg gggatttgtc tttgactgag aaaagagggg
30 181 ctacgtaaaag gctggaccct ggaccccaga ggctgcgggtg gagcaccctg aggcagttcg
241 gcagcttcat cgggagttcc tcagagctgg atcgaacgtc atgcagacct tcactttcta
301 tgcaagttag gacaagctgg aaaaccgagg gaactacgtg gcagagaaga tatctgggca
361 gaaggtcaat gaagctgctt gtgacattgc acggcaagtt gctgacgaag gggatgcatt
421 ggttgcagga ggtgtgagtc agacaccttc ctacctcagc tgcaagagtg agacgggaagt
35 481 taaaaagata tttcaccaac agcttgagggt cttcatgaag aagaatgtgg acttctcat
541 tgcagagtat tttgaacatg ttgaagaagc cgtgtgggca gtcgaggcct taaaaacatc
601 cgggaagcct atagcggcta ccatgtgcat cggacctgaa ggagatctac atggcgtgtc
661 tcctggagag tgccgagtg gtttggtaaa agcaggtgcc gccattgtcg gtgtgaactg
721 ccacttcgac ccagcacca gcttgagac aataaagctc atgaaggagg gtctggaagc
40 781 agctcggctg aaggcttact tgatgagcca cgccctggcc taccacaccc ctgactgtgg
841 caaacaggga tttattgatc tcccagaatt cccctttgga ttggaacca gatttgccac
901 cagatgggat attcaaaaat acgcacagaga ggcctacaac ctgggggtca ggtacattgg
961 cggctgctgc ggatttgagc cctaccacat cagggccatt gcagaggagc tcgccccaga
1021 aaggggattt ttaccaccag cttcagaaaa acatggcagc tggggaagtg gtttggacat
45 1081 gcacacaaa ccttgatca gggcaagggc caggaaagaa tactggcaga atcttcgaat
1141 agcttcgggc agaccgtaca atccttcgat gtccaagccg gatgcttggg gattgacgaa
1201 aggggcagca gagctgatgc agcagaagga agccaccact gagcagcagc tgagagcgct
1261 cttcgaaaaa caaaaattca aatccgcaca gtagccacag gccagcggtt cggggcgaat
1321 tcctccaggt ccgggccaca gtgtgcaccc ggaaggagaa ggcattctta aaccagcggt

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1381  tgtgttgatg  ccggccttaca  cctgtgattg  gtgctagtta  gacaaaatgg  agtcacagat
1441  agcatttcac  agttacaaaa  ctacgcttta  gaattttacc  tagaaggaag  aaaggagaag
1501  tccacagtaa  atcctgaaca  catttcctac  gtgcctgtcg  cattacaggc  gcacaggagt
1561  cactgcagcg  aagagaaagt  caccgcagct  caatctcatt  tcagataggg  ggataggaca
5  1621  ccacctccac  gagtgacata  gaaccattca  gggaccgtat  cataagtga  acagcaacca
1681  tctatatcta  agatgcttcc  caagtggatt  ccaagatctt  ttgagcagga  cccttaggca
1741  gaaacaacac  acaccagccc  tgtaaaactt  aacagataac  tgatccattc  tgtaattctg
1801  taatctctgt  tctgactgct  tccattccat  ttcattaata  aaaacatgcc  ggttgaaaac
1861  cttcaaaaaa  aaaaaaaaaa  aaaa

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Principle components analysis was used to generate three eigenvectors used to transform the original expression level data matrix, as shown in Table 6 below. Eigenvector 1 values represent NSAIDs associated with hepatotoxicity involving hepatocellular damage, Eigenvector 2 values represent NSAIDs associated with hepatotoxicity involving cholestasis, and Eigenvector 3 values represent NSAIDs associated with hepatotoxicity involving elevated transaminase level.

Table 6: Transform Eigenvectors for Hepatotoxicity by Injury Type

Gene	Eigenvector 1	Eigenvector 2	Eigenvector3
INJURYMARKER1	58.7	0.325	-15.2
INJURYMARKER2	20.5	-3.23	3.01
INJURYMARKER3	-16.9	-6.52	-2.09
INJURYMARKER4	-10.3	0.351	-1.45
INJURYMARKER5	-7.59	-0.152	-0.310
INJURYMARKER6	11.4	-2.69	2.49
INJURYMARKER7	-16.0	-1.57	8.71
INJURYMARKER8	-11.6	1.13	5.36
INJURYMARKER9	-11.0	-0.351	0.078
INJURYMARKER10	7.55	0.618	4.65
% of variation explained	99.0	0.7	0.3

These eigenvectors may be used to transform the expression levels of INJURYMARKERS 1-10 ("INJURYMARKERS") in response to a given drug, in order to predict that drug's hepatotoxicity injury type. For example, expression levels of INJURYMARKERS correlating with Eigenvector 1 indicates that the test drug has a risk of hepatotoxicity involving hepatocellular damage. Alternatively, a drug's INJURYMARKERS expression profile can be generated simultaneously with the above-described training set (or an equivalent set) run in parallel with the test drug, and expression levels associated with the test drug directly compared to those of the training set.

GENERAL METHODS

The RISKMARKER (*i.e.* RISKMARKERS 1-8) and INJURYMARKER (*i.e.* INJURYMARKERS 1-10) nucleic acids and encoded polypeptides can be identified using the information provided above. In some embodiments, the RISKMARKER or INJURYMARKER
5 nucleic acids and polypeptide correspond to nucleic acids or polypeptides which include the various sequences (referenced by SEQ ID NOs) disclosed for each RISKMARKER or INJURYMARKER polypeptide.

In its various aspects and embodiments, the invention includes providing a test cell population which includes at least one cell that is capable of expressing one or more of the
10 sequences RISKMARKER 1-8 or INJURYMARKER 1-10. By “capable of expressing” is meant that the gene is present in an intact form in the cell and can be expressed. Expression of one, some, or all of the RISKMARKER or INJURYMARKER sequences is then detected, if present, and, preferably, measured. Using sequence information provided by the database entries for the known sequences, or the sequence information for the newly described sequences,
15 expression of the RISKMARKER or INJURYMARKER sequences can be detected (if present) and measured using techniques well known to one of ordinary skill in the art. For example, sequences within the sequence database entries corresponding to RISKMARKER or INJURYMARKER sequences, or within the sequences disclosed herein, can be used to construct probes for detecting RISKMARKER or INJURYMARKER RNA sequences in, *e.g.*, northern
20 blot hybridization analyses or methods which specifically, and, preferably, quantitatively amplify specific nucleic acid sequences. As another example, the sequences can be used to construct primers for specifically amplifying the RISKMARKER or INJURYMARKER sequences in, *e.g.*, amplification-based detection methods such as reverse-transcription based polymerase chain reaction. When alterations in gene expression are associated with gene amplification or deletion,
25 sequence comparisons in test and reference populations can be made by comparing relative amounts of the examined DNA sequences in the test and reference cell populations.

Expression can be also measured at the protein level, *i.e.*, by measuring the levels of polypeptides encoded by the gene products described herein. Such methods are well known in the art and include, *e.g.*, immunoassays based on antibodies to proteins encoded by the genes.

Expression level of one or more of the RISKMARKER or INJURYMARKER sequences in the test cell population, *e.g.* rat hepatocytes, is then compared to expression levels of the sequences in one or more cells from a reference cell population. Expression of sequences in test and control populations of cells can be compared using any art-recognized method for comparing expression of nucleic acid sequences. For example, expression can be compared using GENE CALLING[®] methods as described in US Patent No. 5,871,697 and in Shimkets et al., Nat. Biotechnol. 17:798-803.

In various embodiments, the expression of one or more sequences which are markers of hepatotoxicity risk, *i.e.* RISKMARKERS 1-8, is compared. In other embodiments, the expression of one or more sequences which are markers of hepatotoxicity injury type, *i.e.* INJURYMARKERS, is compared. In further embodiments, expression of one or more RISKMARKERS and INJURYMARKERS may be compared to predict both hepatotoxicity risk and type of hepatotoxicity injury.

In various embodiments, the expression of 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, or all of the sequences represented by RISKMARKER 1-8 and INJURYMARKER 1-10 are measured. If desired, expression of these sequences can be measured along with other sequences whose expression is known to be altered according to one of the herein described parameters or conditions.

The reference cell population includes one or more cells for which the compared parameter is known. The compared parameter can be, *e.g.*, hepatotoxic agent expression status. By "hepatotoxic agent expression status" is meant that it is known whether the reference cell has had contact with a hepatotoxic agent. An example of a hepatotoxic agent is, *e.g.*, a thiazolidinedione such as troglitazone. Whether or not comparison of the gene expression profile in the test cell population to the reference cell population reveals the presence, or degree, of the measured parameter depends on the composition of the reference cell population. For example, if the reference cell population is composed of cells that have not been treated with a known hepatotoxic agent, a similar gene expression level in the test cell population and a reference cell population indicates the test agent is not a hepatotoxic agent. Conversely, if the reference cell population is made up of cells that have been treated with a hepatotoxic agent, a similar gene

expression profile between the test cell population and the reference cell population indicates the test agent is a hepatotoxic agent.

In various embodiments, a RISKMARKER or INJURYMARKER sequence in a test cell population is considered comparable in expression level to the expression level of the

5 RISKMARKER or INJURYMARKER sequence if its expression level varies within a factor of 2.0, 1.5, or 1.0 fold to the level of the RISKMARKER or INJURYMARKER transcript in the reference cell population. In various embodiments, a RISKMARKER or INJURYMARKER sequence in a test cell population can be considered altered in levels of expression if its expression level varies from the reference cell population by more than 1.0, 1.5, 2.0 or more fold
10 from the expression level of the corresponding RISKMARKER or INJURYMARKER sequence in the reference cell population.

Alternatively, the absolute expression level matrix of the 8 RISKMARKER and/or 10 INJURYMARKER fragments in a test cell can be transformed using the principal component eigenvectors described above, or similar eigenvalues generated from parallel dosed members of the training set as internal controls. The expression eigenvalues for the test cell can then be
15 compared to the training set eigenvalues described herein, or a parallel-run training set, if any.

The RISKMARKER expression level combination is considered similar to Low Risk idiosyncratic NSAIDS (several of which have been withdrawn), if the test drug's expression profile is within the 95% confidence interval (CI) of the centroid of that risk class. See Table 4.
20 The test drug is considered Very Low Risk idiosyncratic if the transformed expression profile falls within the 95% CI of the centroid of that class. The test drug is considered Overdose Risk if the expression profile falls within the 95% CI of the centroid of that class. If the compound fails to associate with any of these compounds it will be categorized as unclassifiable.

Similarly, the INJURYMARKER expression level combination is considered indicative
25 of hepatocellular damage induced by idiosyncratic NSAIDS, if the test drug's expression profile is within the 95% confidence interval (CI) of the centroid of that injury type. See Table 6. The test drug is considered to induce idiosyncratic cholestasis if the transformed expression profile falls within the 95% CI of the centroid of that injury type. The test drug is considered to induce elevated transaminase level if the expression profile falls within the 95% CI of the centroid of

that class. If the compound fails to associate with any of these compounds it will be categorized as unclassifiable.

If desired, comparison of differentially expressed sequences between a test cell population and a reference cell population can be done with respect to a control nucleic acid whose expression is independent of the parameter or condition being measured. Expression levels of the control nucleic acid in the test and reference nucleic acid can be used to normalize signal levels in the compared populations.

In some embodiments, the test cell population is compared to multiple reference cell populations. Each of the multiple reference populations may differ in the known parameter. Thus, a test cell population may be compared to a first reference cell population known to have been exposed to a hepatotoxic agent, as well as a second reference population known have not been exposed to a hepatotoxic agent.

The test cell population that is exposed to, *i.e.*, contacted with, the test hepatotoxic agent can be any number of cells, *i.e.*, one or more cells, and can be provided *in vitro*, *in vivo*, or *ex vivo*.

In other embodiments, the test cell population can be divided into two or more subpopulations. The subpopulations can be created by dividing the first population of cells to create as identical a subpopulation as possible. This will be suitable, in, for example, *in vitro* or *ex vivo* screening methods. In some embodiments, various sub populations can be exposed to a control agent, and/or a test agent, multiple test agents, or, *e.g.*, varying dosages of one or multiple test agents administered together, or in various combinations.

Preferably, cells in the reference cell population are derived from a tissue type as similar as possible to test cell, *e.g.*, liver tissue. In some embodiments, the control cell is derived from the same subject as the test cell, *e.g.*, from a region proximal to the region of origin of the test cell. In other embodiments, the reference cell population is derived from a plurality of cells. For example, the reference cell population can be a database of expression patterns from previously tested cells for which one of the herein-described parameters or conditions (hepatotoxic agent expression status is known.

The test agent can be a compound not previously described or can be a previously known compound but which is not known to be a hepatotoxic agent.

The subject is preferably a mammal. The mammal can be, *e.g.*, a human, non-human primate, mouse, rat, dog, cat, horse, or cow.

5 SCREENING FOR TOXIC AGENTS

In one aspect, the invention provides a method of identifying toxic agents, *e.g.*, hepatotoxic agents. The hepatotoxic agent can be identified by providing a cell population that includes cells capable of expressing one or more nucleic acid sequences homologous to those of RISKMARKER 1-8 or INJURYMARKER 1-10. The sequences need not be identical to sequences including RISKMARKER or INJURYMARKER nucleic acid sequences, as long as the sequence is sufficiently similar that specific hybridization can be detected. Preferably, the cell includes sequences that are identical, or nearly identical to those identifying the RISKMARKER or INJURYMARKER nucleic acids described herein.

Expression of the nucleic acid sequences in the test cell population is then compared to the expression of the nucleic acid sequences in a reference cell population, which is a cell population that has not been exposed to the test agent, or, in some embodiments, a cell population exposed the test agent. Comparison can be performed on test and reference samples measured concurrently or at temporally distinct times. An example of the latter is the use of compiled expression information, *e.g.*, a sequence database, which assembles information about expression levels of known sequences following administration of various agents. For example, alteration of expression levels following administration of test agent can be compared to the expression changes observed in the nucleic acid sequences following administration of a control agent, *e.g.* a NSAID such as ketoprofen.

An alteration in expression of the nucleic acid sequence in the test cell population compared to the expression of the nucleic acid sequence in the reference cell population that has not been exposed to the test agent indicates the test agent is a hepatotoxic agent.

The invention also includes a hepatotoxic agent identified according to this screening method.

In some embodiments of the method of the invention, the test agent is an idiosyncratic hepatotoxic agent, *e.g.* a NSAID, and the reference agent is also a NSAID. As described above, RISKMARKER (*e.g.* RISKMARKERS 1-8) expression level patterns can be used to predict the level of hepatotoxicity risk (*i.e.* low, very low, or overdose) associated with a given test agent, *e.g.* a NSAID. In one embodiment, the reference NSAID (*i.e.* used with the reference cell population) is a NSAID classified as having a low risk of hepatotoxicity. The test agent is then identified as having a low risk of hepatotoxicity if no qualitative difference in expression levels is identified in comparison to expression levels in the reference population exposed to a low risk NSAID. In certain embodiments, the low risk NSAID is Benoxaprofen, Bromfenac, Diclofenac, Phenylbutazone, or Sulindac. In another embodiment, the reference NSAID is a NSAID classified as having a very low risk of hepatotoxicity. The test agent is then identified as having a very low risk of hepatotoxicity if no qualitative difference in expression levels is identified in comparison to expression levels in the reference population exposed to a very low risk NSAID. In certain embodiments, the very low risk NSAID is Etodolac, Fenoprofen, Flurbiprofen, Ibuprofen, Indomethacin, Ketoprofen, Meclofenamate, Mefenamic Acid, Nabumetone, Naproxen, Oxaprozin, Piroxicam, Suprofen, Tenoxicam, Tolmentin, or Zomepirac. In still another embodiment, the reference NSAID is a NSAID classified as having an overdose risk of hepatotoxicity. The test agent is then identified as having an overdose risk of hepatotoxicity if no qualitative difference in expression levels is identified in comparison to expression levels in the reference population exposed to an overdose risk NSAID. In certain embodiments, the overdose risk NSAID is Acetaminophen, Acetylsalicylic acid, or Phenacetin. In some embodiments, the difference in expression levels is determined by comparing expression transformation eigenvectors (for risk class) for the test cell and reference cell populations, as described above.

As also described above, INJURYMARKER (*e.g.* INJURYMARKERS 1-10) expression level patterns can be used to predict the type of hepatotoxicity injury (*i.e.* hepatocellular damage, cholestasis, or elevated transaminase level) associated with a given test agent, *e.g.* a NSAID. In some embodiments, the reference NSAID is a NSAID classified as inducing hepatocellular damage. The test agent is then identified as likely to induce hepatocellular damage if no qualitative difference in expression levels is identified in comparison to expression levels in the reference population exposed to a NSAID which induces hepatocellular damage. In certain embodiments, the hepatocellular damage inducing NSAID is Acetaminophen, Flurbiprofen, or

Ketoprofen. In another embodiment, the reference NSAID is a NSAID classified as inducing cholestasis. The test agent is then identified as likely to induce cholestasis if no qualitative difference in expression levels is identified in comparison to expression levels in the reference population exposed to a NSAID which induces cholestasis. In certain embodiments, the cholestatis-inducing NSAID is Benoxaprofen, Nabumetone, or Sulindac. In yet another embodiment, the reference NSAID is a NSAID classified as inducing elevated transaminase level. The test agent is then identified as likely to induce elevated transaminase level if no qualitative difference in expression levels is identified as compared to expression levels in the reference population exposed to a NSAID which induces elevated transaminase levels. In certain embodiments, the elevated transaminase level inducing NSAID is Zomepirac, Mefenamic acid, or Tenoxicam. In some embodiments, the difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations, as described above.

ASSESSING TOXICITY OF A TOXIC AGENT IN A SUBJECT

The differentially expressed RISKMARKER or INJURYMARKER sequences identified herein also allow for the hepatotoxicity of a hepatotoxic agent to be determined or monitored. In this method, a test cell population from a subject is exposed to a test agent, *i.e.* a hepatotoxic agent. If desired, test cell populations can be taken from the subject at various time points before, during, or after exposure to the test agent. Expression of one or more of the RISKMARKER or INJURYMARKER sequences in the cell population is then measured and compared to a reference cell population which includes cells whose hepatotoxic agent expression status is known. Preferably, the reference cells not been exposed to the test agent.

If the reference cell population contains no cells exposed to the treatment, a similarity in expression between RISKMARKER or INJURYMARKER sequences in the test cell population and the reference cell population indicates that the treatment is non-hepatotoxic. However, a difference in expression between RISKMARKER or INJURYMARKER sequences in the test population and this reference cell population indicates the treatment is hepatotoxic.

By “hepatotoxicity” is meant that the agent is damaging or destructive to liver when administered to a subject leads to liver damage.

As described in detail above, RISKMARKER expression patterns can be used to predict the level of hepatotoxicity risk (*e.g.* low risk, very low risk, overdose risk) associated with a test agent or drug, by comparison to RISKMARKER expression levels for reference drugs, *e.g.* NSAIDs, with a given classification of risk (*e.g.* very low risk). Similarly, INJURYMARKER expression patterns can be used to predict the type of hepatotoxicity damage (*e.g.* hepatocellular damage, cholestasis, elevated transaminase level) associated with a test agent or drug, by comparison to INJURYMARKER expression levels for reference drugs, *e.g.* NSAIDs, which induce a given type of hepatotoxic damage (*e.g.* cholestasis).

RISKMARKER NUCLEIC ACIDS

Also provided in the invention are novel nucleic acid comprising a nucleic acid sequence selected from the group consisting of RISKMARKER 1, and RISKMARKERS 6-8, or their complements, as well as vectors and cells including these nucleic acids.

Thus, one aspect of the invention pertains to isolated RISKMARKER nucleic acid molecules that encode RISKMARKER proteins or biologically active portions thereof. Also included are nucleic acid fragments sufficient for use as hybridization probes to identify RISKMARKER-encoding nucleic acids (*e.g.*, RISKMARKER mRNA) and fragments for use as polymerase chain reaction (PCR) primers for the amplification or mutation of RISKMARKER nucleic acid molecules. As used herein, the term “nucleic acid molecule” is intended to include DNA molecules (*e.g.*, cDNA or genomic DNA), RNA molecules (*e.g.*, mRNA), analogs of the DNA or RNA generated using nucleotide analogs, and derivatives, fragments and homologs thereof. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

“Probes” refer to nucleic acid sequences of variable length, preferably between at least about 10 nucleotides (nt) or as many as about, *e.g.*, 6,000 nt, depending on use. Probes are used in the detection of identical, similar, or complementary nucleic acid sequences. Longer length probes are usually obtained from a natural or recombinant source, are highly specific and much slower to hybridize than oligomers. Probes may be single- or double-stranded and designed to have specificity in PCR, membrane-based hybridization technologies, or ELISA-like technologies.

An "isolated" nucleic acid molecule is one that is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. Examples of isolated nucleic acid molecules include, but are not limited to, recombinant DNA molecules contained in a vector, recombinant DNA molecules maintained in a heterologous host cell, partially or substantially purified nucleic acid molecules, and synthetic DNA or RNA molecules. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated RISKMARKER nucleic acid molecule can contain less than about 50 kb, 25 kb, 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material or culture medium when produced by recombinant techniques, or of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, *e.g.*, a nucleic acid molecule having the nucleotide sequence of any of RISKMARKER 1, or RISKMARKER 6-8, or a complement of any of these nucleotide sequences, can be isolated using standard molecular biology techniques and the sequence information provided herein. Using all or a portion of these nucleic acid sequences as a hybridization probe, RISKMARKER or INJURYMARKER nucleic acid sequences can be isolated using standard hybridization and cloning techniques (*e.g.*, as described in Sambrook *et al.*, eds., *Molecular Cloning: A Laboratory Manual* 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989; and Ausubel, *et al.*, eds., *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, NY, 1993.)

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to RISKMARKER nucleotide sequences can be prepared by standard synthetic techniques, *e.g.*, using an automated DNA synthesizer.

As used herein, the term “oligonucleotide” refers to a series of linked nucleotide residues, which oligonucleotide has a sufficient number of nucleotide bases to be used in a PCR reaction. A short oligonucleotide sequence may be based on, or designed from, a genomic or cDNA sequence and is used to amplify, confirm, or reveal the presence of an identical, similar or
 5 complementary DNA or RNA in a particular cell or tissue. Oligonucleotides comprise portions of a nucleic acid sequence having at least about 10 nt and as many as 50 nt, preferably about 15 nt to 30 nt. They may be chemically synthesized and may be used as probes.

In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule that is a complement of the nucleotide sequence shown in RISKMARKER
 10 1, or RISKMARKER 6-8. In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule that is a complement of the nucleotide sequence shown in any of these sequences, or a portion of any of these nucleotide sequences. A nucleic acid molecule that is complementary to the nucleotide sequence shown in RISKMARKER 1, or RISKMARKER 6-8 is one that is sufficiently complementary to the nucleotide sequence shown,
 15 such that it can hydrogen bond with little or no mismatches to the nucleotide sequences shown, thereby forming a stable duplex.

As used herein, the term “complementary” refers to Watson-Crick or Hoogsteen base pairing between nucleotides units of a nucleic acid molecule, and the term “binding” means the physical or chemical interaction between two polypeptides or compounds or associated
 20 polypeptides or compounds or combinations thereof. Binding includes ionic, non-ionic, Von der Waals, hydrophobic interactions, etc. A physical interaction can be either direct or indirect. Indirect interactions may be through or due to the effects of another polypeptide or compound. Direct binding refers to interactions that do not take place through, or due to, the effect of another polypeptide or compound, but instead are without other substantial chemical
 25 intermediates.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of RISKMARKER 1, or RISKMARKER 6-8, *e.g.*, a fragment that can be used as a probe or primer or a fragment encoding a biologically active portion of
 30 RISKMARKER. Fragments provided herein are defined as sequences of at least 6 (contiguous) nucleic acids or at least 4 (contiguous) amino acids, a length sufficient to allow for specific

hybridization in the case of nucleic acids or for specific recognition of an epitope in the case of amino acids, respectively, and are at most some portion less than a full length sequence.

Fragments may be derived from any contiguous portion of a nucleic acid or amino acid sequence of choice. Derivatives are nucleic acid sequences or amino acid sequences formed from the native compounds either directly or by modification or partial substitution. Analogs are nucleic acid sequences or amino acid sequences that have a structure similar to, but not identical to, the native compound but differs from it in respect to certain components or side chains. Analogs may be synthetic or from a different evolutionary origin and may have a similar or opposite metabolic activity compared to wild type.

Derivatives and analogs may be full length or other than full length, if the derivative or analog contains a modified nucleic acid or amino acid, as described below. Derivatives or analogs of the nucleic acids or proteins of the invention include, but are not limited to, molecules comprising regions that are substantially homologous to the nucleic acids or proteins of the invention, in various embodiments, by at least about 45%, 50%, 70%, 80%, 95%, 98%, or even 99% identity (with a preferred identity of 80-99%) over a nucleic acid or amino acid sequence of identical size or when compared to an aligned sequence in which the alignment is done by a computer homology program known in the art, or whose encoding nucleic acid is capable of hybridizing to the complement of a sequence encoding the aforementioned proteins under stringent, moderately stringent, or low stringent conditions. See *e.g.* Ausubel, *et al.*, Current Protocols in Molecular Biology, John Wiley & Sons, New York, NY, 1993, and below. An exemplary program is the Gap program (Wisconsin Sequence Analysis Package, Version 8 for UNIX, Genetics Computer Group, University Research Park, Madison, WI) using the default settings, which uses the algorithm of Smith and Waterman (Adv. Appl. Math., 1981, 2: 482-489, which is incorporated herein by reference in its entirety).

A “homologous nucleic acid sequence” or “homologous amino acid sequence,” or variations thereof, refer to sequences characterized by a homology at the nucleotide level or amino acid level as discussed above. Homologous nucleotide sequences encode those sequences coding for isoforms of a RISKMARKER polypeptide. Isoforms can be expressed in different tissues of the same organism as a result of, for example, alternative splicing of RNA.

Alternatively, isoforms can be encoded by different genes. In the present invention, homologous nucleotide sequences include nucleotide sequences encoding for a RISKMARKER polypeptide

of species other than humans, including, but not limited to, mammals, and thus can include, *e.g.*, mouse, rat, rabbit, dog, cat, cow, horse, and other organisms. Homologous nucleotide sequences also include, but are not limited to, naturally occurring allelic variations and mutations of the nucleotide sequences set forth herein. A homologous nucleotide sequence does not, however, include the nucleotide sequence encoding a human RISKMARKER protein. Homologous nucleic acid sequences include those nucleic acid sequences that encode conservative amino acid substitutions (see below) in a RISKMARKER polypeptide, as well as a polypeptide having a RISKMARKER activity. A homologous amino acid sequence does not encode the amino acid sequence of a human RISKMARKER polypeptide.

The nucleotide sequence determined from the cloning of human RISKMARKER genes allows for the generation of probes and primers designed for use in identifying and/or cloning RISKMARKER homologues in other cell types, *e.g.*, from other tissues, as well as RISKMARKER homologues from other mammals. The probe/primer typically comprises a substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12, 25, 50, 100, 150, 200, 250, 300, 350 or 400 consecutive sense strand nucleotide sequence of a nucleic acid comprising a RISKMARKER sequence, or an anti-sense strand nucleotide sequence of a nucleic acid comprising a RISKMARKER sequence, or of a naturally occurring mutant of these sequences.

Probes based on human RISKMARKER nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In various embodiments, the probe further comprises a label group attached thereto, *e.g.*, the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissue which misexpress a RISKMARKER protein, such as by measuring a level of a RISKMARKER-encoding nucleic acid in a sample of cells from a subject *e.g.*, detecting RISKMARKER mRNA levels or determining whether a genomic RISKMARKER gene has been mutated or deleted.

“A polypeptide having a biologically active portion of RISKMARKER” refers to polypeptides exhibiting activity similar, but not necessarily identical to, an activity of a polypeptide of the present invention, including mature forms, as measured in a particular

biological assay, with or without dose dependency. A nucleic acid fragment encoding a "biologically active portion of RISKMARKER" can be prepared by isolating a portion of RISKMARKER 1, or RISKMARKER 6-8, that encodes a polypeptide having a RISKMARKER biological activity, expressing the encoded portion of RISKMARKER protein (*e.g.*, by recombinant expression *in vitro*) and assessing the activity of the encoded portion of RISKMARKER. For example, a nucleic acid fragment encoding a biologically active portion of a RISKMARKER polypeptide can optionally include an ATP-binding domain. In another embodiment, a nucleic acid fragment encoding a biologically active portion of RISKMARKER includes one or more regions.

RISKMARKER AND INJURYMARKER VARIANTS

The invention further encompasses nucleic acid molecules that differ from the disclosed or referenced RISKMARKER or INJURYMARKER nucleotide sequences due to degeneracy of the genetic code. These nucleic acids thus encode the same RISKMARKER or INJURYMARKER protein as that encoded by nucleotide sequence comprising a RISKMARKER or INJURYMARKER nucleic acid as shown in, *e.g.*, RISKMARKER 1-8 INJURYMARKER 1-10.

In addition to the rat RISKMARKER or INJURYMARKER nucleotide sequence shown in RISKMARKER or INJURYMARKER 1, and RISKMARKER or INJURYMARKER 6-8, it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of a RISKMARKER or INJURYMARKER polypeptide may exist within a population (*e.g.*, the human population). Such genetic polymorphism in the RISKMARKER or INJURYMARKER gene may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame encoding a RISKMARKER or INJURYMARKER protein, preferably a mammalian RISKMARKER or INJURYMARKER protein. Such natural allelic variations can typically result in 1-5% variance in the nucleotide sequence of the RISKMARKER or INJURYMARKER gene. Any and all such nucleotide variations and resulting amino acid polymorphisms in RISKMARKER or INJURYMARKER

that are the result of natural allelic variation and that do not alter the functional activity of RISKMARKER or INJURYMARKER are intended to be within the scope of the invention.

Moreover, nucleic acid molecules encoding RISKMARKER or INJURYMARKER proteins from other species, and thus that have a nucleotide sequence that differs from the human sequence of RISKMARKER OR INJURYMARKER, are intended to be within the scope of the invention. Nucleic acid molecules corresponding to natural allelic variants and homologues of the RISKMARKER or INJURYMARKER DNAs of the invention can be isolated based on their homology to the human RISKMARKER or INJURYMARKER nucleic acids disclosed herein using the human cDNAs, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. For example, a soluble human RISKMARKER or INJURYMARKER DNA can be isolated based on its homology to human membrane-bound RISKMARKER or INJURYMARKER. Likewise, a membrane-bound human RISKMARKER or INJURYMARKER DNA can be isolated based on its homology to soluble human RISKMARKER or INJURYMARKER.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 6 nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of RISKMARKER 1, or RISKMARKER 6-8. In another embodiment, the nucleic acid is at least 10, 25, 50, 100, 250 or 500 nucleotides in length. In another embodiment, an isolated nucleic acid molecule of the invention hybridizes to the coding region. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% homologous to each other typically remain hybridized to each other.

Homologs (*i.e.*, nucleic acids encoding RISKMARKER or INJURYMARKER proteins derived from species other than human) or other related sequences (*e.g.*, paralogs) can be obtained by low, moderate or high stringency hybridization with all or a portion of the particular human sequence as a probe using methods well known in the art for nucleic acid hybridization and cloning.

As used herein, the phrase "stringent hybridization conditions" refers to conditions under which a probe, primer or oligonucleotide will hybridize to its target sequence, but to no other sequences. Stringent conditions are sequence-dependent and will be different in different

circumstances. Longer sequences hybridize specifically at higher temperatures than shorter sequences. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present at excess, at T_m, 50% of the probes are occupied at equilibrium. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes, primers or oligonucleotides (e.g., 10 nt to 50 nt) and at least about 60°C for longer probes, primers and oligonucleotides. Stringent conditions may also be achieved with the addition of destabilizing agents, such as formamide.

Stringent conditions are known to those skilled in the art and can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. Preferably, the conditions are such that sequences at least about 65%, 70%, 75%, 85%, 90%, 95%, 98%, or 99% homologous to each other typically remain hybridized to each other. A non-limiting example of stringent hybridization conditions is hybridization in a high salt buffer comprising 6X SSC, 50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.02% BSA, and 500 mg/ml denatured salmon sperm DNA at 65°C. This hybridization is followed by one or more washes in 0.2X SSC, 0.01% BSA at 50°C. An isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of RISKMARKER 1, or RISKMARKER 6-8 corresponds to a naturally occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein).

In a second embodiment, a nucleic acid sequence that is hybridizable to the nucleic acid molecule comprising the nucleotide sequence of RISKMARKER 1, or RISKMARKER 6-8, or fragments, analogs or derivatives thereof, under conditions of moderate stringency is provided. A non-limiting example of moderate stringency hybridization conditions are hybridization in 6X SSC, 5X Denhardt's solution, 0.5% SDS and 100 mg/ml denatured salmon sperm DNA at 55°C, followed by one or more washes in 1X SSC, 0.1% SDS at 37°C. Other conditions of moderate stringency that may be used are well known in the art. See, e.g., Ausubel *et al.* (eds.), 1993,

Current Protocols in Molecular Biology, John Wiley & Sons, NY, and Kriegler, 1990, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY.

In a third embodiment, a nucleic acid that is hybridizable to the nucleic acid molecule comprising the nucleotide sequence of RISKMARKER 1, or RISKMARKER 6-8, or fragments, analogs or derivatives thereof, under conditions of low stringency, is provided. A non-limiting example of low stringency hybridization conditions are hybridization in 35% formamide, 5X SSC, 50 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.2% BSA, 100 mg/ml denatured salmon sperm DNA, 10% (wt/vol) dextran sulfate at 40°C, followed by one or more washes in 2X SSC, 25 mM Tris-HCl (pH 7.4), 5 mM EDTA, and 0.1% SDS at 50°C.

Other conditions of low stringency that may be used are well known in the art (*e.g.*, as employed for cross-species hybridizations). See, *e.g.*, Ausubel *et al.* (eds.), 1993, Current Protocols in Molecular Biology, John Wiley & Sons, NY, and Kriegler, 1990, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY; Shilo *et al.*, 1981, *Proc Natl Acad Sci USA* 78: 6789-6792.

15 CONSERVATIVE MUTATIONS

In addition to naturally-occurring allelic variants of the RISKMARKER sequence that may exist in the population, the skilled artisan will further appreciate that changes can be introduced into an RISKMARKER nucleic acid or directly into an RISKMARKER polypeptide sequence without altering the functional ability of the RISKMARKER protein. In some embodiments, the nucleotide sequence of RISKMARKER 1, or RISKMARKER 6-8 will be altered, thereby leading to changes in the amino acid sequence of the encoded RISKMARKER protein. For example, nucleotide substitutions that result in amino acid substitutions at various "non-essential" amino acid residues can be made in the sequence of RISKMARKER 1, or RISKMARKER 6-8. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of RISKMARKER without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the RISKMARKER proteins of the present invention, are predicted to be particularly unamenable to alteration.

In addition, amino acid residues that are conserved among family members of the RISKMARKER proteins of the present invention, are also predicted to be particularly unamenable to alteration. As such, these conserved domains are not likely to be amenable to mutation. Other amino acid residues, however, (*e.g.*, those that are not conserved or only
 5 semi-conserved among members of the RISKMARKER proteins) may not be essential for activity and thus are likely to be amenable to alteration.

Another aspect of the invention pertains to nucleic acid molecules encoding RISKMARKER proteins that contain changes in amino acid residues that are not essential for activity. Such RISKMARKER proteins differ in amino acid sequence from the amino acid
 10 sequences of polypeptides encoded by nucleic acids containing RISKMARKER 1, or RISKMARKER 6-8, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 45% homologous, more preferably 60%, and still more preferably at least about 70%, 80%, 90%, 95%, 98%, and most preferably at least about 99%
 15 homologous to the amino acid sequence of the amino acid sequences of polypeptides encoded by nucleic acids comprising RISKMARKER 1, or RISKMARKER 6-8.

An isolated nucleic acid molecule encoding a RISKMARKER protein homologous to can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of a nucleic acid comprising RISKMARKER 1, or RISKMARKER 6-8,
 20 such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein.

Mutations can be introduced into a nucleic acid comprising RISKMARKER 1, or RISKMARKER 6-8 by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one
 25 or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (*e.g.*, lysine, arginine, histidine), acidic side chains (*e.g.*, aspartic acid, glutamic acid), uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (*e.g.*, alanine,
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valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (*e.g.*, threonine, valine, isoleucine) and aromatic side chains (*e.g.*, tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in RISKMARKER is replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of a RISKMARKER coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for RISKMARKER biological activity to identify mutants that retain activity. Following mutagenesis of the nucleic acid, the encoded protein can be expressed by any recombinant technology known in the art and the activity of the protein can be determined.

In one embodiment, a mutant RISKMARKER protein can be assayed for (1) the ability to form protein:protein interactions with other RISKMARKER proteins, other cell-surface proteins, or biologically active portions thereof, (2) complex formation between a mutant RISKMARKER protein and a RISKMARKER ligand; (3) the ability of a mutant RISKMARKER protein to bind to an intracellular target protein or biologically active portion thereof; (*e.g.*, avidin proteins); (4) the ability to bind ATP; or (5) the ability to specifically bind a RISKMARKER protein antibody.

In other embodiment, the fragment of the complementary polynucleotide sequence described in claim 1 wherein the fragment of the complementary polynucleotide sequence hybridizes to the first sequence.

In other specific embodiments, the nucleic acid is RNA or DNA. The fragment or the fragment of the complementary polynucleotide sequence described in claim 38, wherein the fragment is between about 10 and about 100 nucleotides in length, *e.g.*, between about 10 and about 90 nucleotides in length, or about 10 and about 75 nucleotides in length, about 10 and about 50 bases in length, about 10 and about 40 bases in length, or about 15 and about 30 bases in length.

ANTI-SENSE

Another aspect of the invention pertains to isolated antisense nucleic acid molecules that are hybridizable to or complementary to the nucleic acid molecule comprising the nucleotide sequence of a RISKMARKER or INJURYMARKER sequence or fragments, analogs or derivatives thereof. An "antisense" nucleic acid comprises a nucleotide sequence that is

complementary to a "sense" nucleic acid encoding a protein, *e.g.*, complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. In specific aspects, antisense nucleic acid molecules are provided that comprise a sequence complementary to at least about 10, 25, 50, 100, 250 or 500 nucleotides or an entire

5 RISKMARKER or INJURYMARKER coding strand, or to only a portion thereof. Nucleic acid molecules encoding fragments, homologs, derivatives and analogs of a RISKMARKER or INJURYMARKER protein, or antisense nucleic acids complementary to a nucleic acid comprising a RISKMARKER or INJURYMARKER nucleic acid sequence are additionally provided.

10 In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding RISKMARKER or INJURYMARKER. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues. In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence
15 encoding RISKMARKER. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (*i.e.*, also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding RISKMARKER or INJURYMARKER disclosed herein, antisense nucleic acids of the invention can be designed according to the rules
20 of Watson and Crick or Hoogsteen base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of RISKMARKER or INJURYMARKER mRNA, but more preferably is an oligonucleotide that is antisense to only a portion of the coding or noncoding region of RISKMARKER or INJURYMARKER mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of
25 RISKMARKER or INJURYMARKER mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis or enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or
30 variously modified nucleotides designed to increase the biological stability of the molecules or to

increase the physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used.

Examples of modified nucleotides that can be used to generate the antisense nucleic acid include: 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding a RISKMARKER or INJURYMARKER protein to thereby inhibit expression of the protein, *e.g.*, by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule that binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention includes direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, *e.g.*, by linking the antisense nucleic acid molecules to peptides or antibodies that bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered

to cells using the vectors described herein. To achieve sufficient intracellular concentrations of antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual α -units, the strands run parallel to each other (Gaultier *et al.* (1987) *Nucleic Acids Res* 15: 6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue *et al.* (1987) *Nucleic Acids Res* 15: 6131-6148) or a chimeric RNA -DNA analogue (Inoue *et al.* (1987) *FEBS Lett* 215: 327-330).

RIBOZYMES AND PNA MOIETIES

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity that are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (*e.g.*, hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to catalytically cleave RISKMARKER or INJURYMARKER mRNA transcripts to thereby inhibit translation of RISKMARKER or INJURYMARKER mRNA. A ribozyme having specificity for a RISKMARKER or INJURYMARKER -encoding nucleic acid can be designed based upon the nucleotide sequence of a RISKMARKER or INJURYMARKER DNA disclosed herein. For example, a derivative of a Tetrahymena L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a RISKMARKER or INJURYMARKER-encoding mRNA. See, *e.g.*, Cech *et al.* U.S. Pat. No. 4,987,071; and Cech *et al.* U.S. Pat. No. 5,116,742. Alternatively, RISKMARKER or INJURYMARKER mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, *e.g.*, Bartel *et al.*, (1993) *Science* 261:1411-1418.

Alternatively, RISKMARKER or INJURYMARKER gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of a RISKMARKER or INJURYMARKER nucleic acid (*e.g.*, the RISKMARKER or INJURYMARKER promoter

and/or enhancers) to form triple helical structures that prevent transcription of the RISKMARKER or INJURYMARKER gene in target cells. See generally, Helene. (1991) *Anticancer Drug Des.* 6: 569-84; Helene. *et al.* (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher (1992) *Bioassays* 14: 807-15.

5 In various embodiments, the nucleic acids of RISKMARKER or INJURYMARKER can be modified at the base moiety, sugar moiety or phosphate backbone to improve, *e.g.*, the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (see Hyrup *et al.* (1996) *Bioorg Med Chem* 4: 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, *e.g.*, DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup *et al.* (1996) above; Perry-O'Keefe *et al.* (1996) *PNAS* 93: 14670-675.

15 PNAs of RISKMARKER or INJURYMARKER can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, *e.g.*, inducing transcription or translation arrest or inhibiting replication. PNAs of RISKMARKER or INJURYMARKER can also be used, *e.g.*, in the analysis of single base pair mutations in a gene by, *e.g.*, PNA directed PCR clamping; as artificial restriction enzymes when used in combination with other enzymes, *e.g.*, S1 nucleases (Hyrup B. (1996) above); or as probes or primers for DNA sequence and hybridization (Hyrup *et al.* (1996), above; Perry-O'Keefe (1996), above).

25 In another embodiment, PNAs of RISKMARKER or INJURYMARKER can be modified, *e.g.*, to enhance their stability or cellular uptake, by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of RISKMARKER or INJURYMARKER can be generated that may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, *e.g.*, RNase H and DNA polymerases, to interact with the DNA portion while the PNA portion would provide

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high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup (1996) above). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996) above and Finn *et al.* (1996) *Nucl Acids Res* 24:

3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry, and modified nucleoside analogs, *e.g.*, 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used between the PNA and the 5' end of DNA (Mag *et al.* (1989) *Nucl Acid Res* 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn *et al.* (1996) above). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment. See, Petersen *et al.* (1975) *Bioorg Med Chem Lett* 5: 1119-11124.

In other embodiments, the oligonucleotide may include other appended groups such as peptides (*e.g.*, for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, *e.g.*, Letsinger *et al.*, 1989, *Proc. Natl. Acad. Sci. U.S.A.* 86:6553-6556; Lemaitre *et al.*, 1987, *Proc. Natl. Acad. Sci.* 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, *e.g.*, PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization triggered cleavage agents (See, *e.g.*, Krol *et al.*, 1988, *BioTechniques* 6:958-976) or intercalating agents. (See, *e.g.*, Zon, 1988, *Pharm. Res.* 5: 539-549). To this end, the oligonucleotide may be conjugated to another molecule, *e.g.*, a peptide, a hybridization triggered cross-linking agent, a transport agent, a hybridization-triggered cleavage agent, etc.

RISKMARKER AND INJURYMARKER POLYPEPTIDES

One aspect of the invention pertains to isolated RISKMARKER or INJURYMARKER proteins, and biologically active portions thereof, or derivatives, fragments, analogs or homologs thereof. Also provided are polypeptide fragments suitable for use as immunogens to raise anti-RISKMARKER or INJURYMARKER antibodies, *e.g.* antibodies against RISKMARKER 1, or RISKMARKER 6-8. In one embodiment, native RISKMARKER or INJURYMARKER proteins can be isolated from cells or tissue sources by an appropriate purification scheme using

standard protein purification techniques. In another embodiment, RISKMARKER or INJURYMARKER proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, a RISKMARKER or INJURYMARKER protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

5 An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the RISKMARKER or INJURYMARKER protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of RISKMARKER or INJURYMARKER protein in
10 which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of RISKMARKER or INJURYMARKER protein having less than about 30% (by dry weight) of non-RISKMARKER or INJURYMARKER protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of
15 non-RISKMARKER or INJURYMARKER protein, still more preferably less than about 10% of non-RISKMARKER or INJURYMARKER protein, and most preferably less than about 5% non-RISKMARKER or INJURYMARKER protein. When the RISKMARKER or INJURYMARKER protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, *i.e.*, culture medium represents less than
20 about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of RISKMARKER or INJURYMARKER protein in which the protein is separated from chemical precursors or other chemicals that are involved in the synthesis of the protein. In
25 one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of RISKMARKER or INJURYMARKER protein having less than about 30% (by dry weight) of chemical precursors or non-RISKMARKER or INJURYMARKER chemicals, more preferably less than about 20% chemical precursors or non-RISKMARKER or INJURYMARKER chemicals, still more preferably less than about 10% chemical precursors or
30 non-RISKMARKER or INJURYMARKER chemicals, and most preferably less than about 5% chemical precursors or non-RISKMARKER or INJURYMARKER chemicals.

Biologically active portions of a RISKMARKER or INJURYMARKER protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the RISKMARKER or INJURYMARKER protein, *e.g.*, the amino acid sequence encoded by a nucleic acid comprising RISKMARKER or INJURYMARKER 1-20 that include fewer amino acids than the full length RISKMARKER or INJURYMARKER proteins, and exhibit at least one activity of a RISKMARKER or INJURYMARKER protein. Typically, biologically active portions comprise a domain or motif with at least one activity of the RISKMARKER or INJURYMARKER protein. A biologically active portion of a RISKMARKER or INJURYMARKER protein can be a polypeptide which is, for example, 10, 25, 50, 100 or more amino acids in length.

A biologically active portion of a RISKMARKER or INJURYMARKER protein of the present invention may contain at least one of the above-identified domains conserved between the RISKMARKER or INJURYMARKER proteins. An alternative biologically active portion of a RISKMARKER or INJURYMARKER protein may contain at least two of the above-identified domains. Another biologically active portion of a RISKMARKER or INJURYMARKER protein may contain at least three of the above-identified domains. Yet another biologically active portion of a RISKMARKER or INJURYMARKER protein of the present invention may contain at least four of the above-identified domains.

Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the functional activities of a native RISKMARKER or INJURYMARKER protein.

In some embodiments, the RISKMARKER or INJURYMARKER protein is substantially homologous to one of these RISKMARKER or INJURYMARKER proteins and retains its the functional activity, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail below.

In specific embodiments, the invention includes an isolated polypeptide comprising an amino acid sequence that is 80% or more identical to the sequence of a polypeptide whose expression is modulated in a mammal to which hepatotoxic agent is administered.

DETERMINING HOMOLOGY BETWEEN TWO OR MORE SEQUENCES

To determine the percent homology of two amino acid sequences or of two nucleic acids, the sequences are aligned for optimal comparison purposes (*e.g.*, gaps can be introduced in the sequence of a first amino acid or nucleic acid sequence for optimal alignment with a second amino or nucleic acid sequence). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are homologous at that position (*i.e.*, as used herein amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity").

The nucleic acid sequence homology may be determined as the degree of identity between two sequences. The homology may be determined using computer programs known in the art, such as GAP software provided in the GCG program package. See *Needleman and Wunsch* 1970 *J Mol Biol* 48: 443-453. Using GCG GAP software with the following settings for nucleic acid sequence comparison: GAP creation penalty of 5.0 and GAP extension penalty of 0.3, the coding region of the analogous nucleic acid sequences referred to above exhibits a degree of identity preferably of at least 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, with the CDS (encoding) part of a DNA sequence comprising RISKMARKER 1, or RISKMARKER 6-8..

The term "sequence identity" refers to the degree to which two polynucleotide or polypeptide sequences are identical on a residue-by-residue basis over a particular region of comparison. The term "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over that region of comparison, determining the number of positions at which the identical nucleic acid base (*e.g.*, A, T, C, G, U, or I, in the case of nucleic acids) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the region of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of sequence identity. The term "substantial identity" as used herein denotes a characteristic of a polynucleotide sequence, wherein the polynucleotide comprises a sequence that has at least 80 percent sequence identity, preferably at least 85 percent identity and often 90 to 95 percent sequence identity, more usually at least 99 percent sequence identity as compared to a reference sequence over a comparison region.

CHIMERIC AND FUSION PROTEINS

The invention also provides RISKMARKER chimeric or fusion proteins. As used herein, an RISKMARKER "chimeric protein" or "fusion protein" comprises an RISKMARKER polypeptide operatively linked to a non-RISKMARKER polypeptide. A "RISKMARKER polypeptide" refers to a polypeptide having an amino acid sequence corresponding to RISKMARKER, whereas a "non-RISKMARKER polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein that is not substantially homologous to the RISKMARKER protein, *e.g.*, a protein that is different from the RISKMARKER protein and that is derived from the same or a different organism. Within an RISKMARKER fusion protein the RISKMARKER polypeptide can correspond to all or a portion of an RISKMARKER protein. In one embodiment, an RISKMARKER fusion protein comprises at least one biologically active portion of an RISKMARKER protein. In another embodiment, an RISKMARKER fusion protein comprises at least two biologically active portions of an RISKMARKER protein. In yet another embodiment, an RISKMARKER fusion protein comprises at least three biologically active portions of an RISKMARKER protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the RISKMARKER polypeptide and the non-RISKMARKER polypeptide are fused in-frame to each other. The non-RISKMARKER polypeptide can be fused to the N-terminus or C-terminus of the RISKMARKER polypeptide.

For example, in one embodiment an RISKMARKER fusion protein comprises an RISKMARKER domain operably linked to the extracellular domain of a second protein. Such fusion proteins can be further utilized in screening assays for compounds which modulate RISKMARKER activity (such assays are described in detail below).

In yet another embodiment, the fusion protein is a GST-RISKMARKER fusion protein in which the RISKMARKER sequences are fused to the C-terminus of the GST (*i.e.*, glutathione S-transferase) sequences. Such fusion proteins can facilitate the purification of recombinant RISKMARKER, *e.g.* RISKMARKER 1, or RISKMARKER 6-8.

In another embodiment, the fusion protein is an RISKMARKER protein containing a heterologous signal sequence at its N-terminus. For example, a native RISKMARKER signal sequence can be removed and replaced with a signal sequence from another protein. In certain

host cells (*e.g.*, mammalian host cells), expression and/or secretion of RISKMARKER can be increased through use of a heterologous signal sequence.

In yet another embodiment, the fusion protein is a RISKMARKER-immunoglobulin fusion protein in which the RISKMARKER sequences comprising one or more domains are fused to sequences derived from a member of the immunoglobulin protein family. The RISKMARKER-immunoglobulin fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject to inhibit an interaction between a RISKMARKER ligand and a RISKMARKER protein on the surface of a cell, to thereby suppress RISKMARKER-mediated signal transduction *in vivo*. The RISKMARKER-immunoglobulin fusion proteins can be used to affect the bioavailability of an RISKMARKER cognate ligand. Inhibition of the RISKMARKER ligand/RISKMARKER interaction may be useful therapeutically for both the treatments of proliferative and differentiative disorders, as well as modulating (*e.g.* promoting or inhibiting) cell survival. Moreover, the RISKMARKER-immunoglobulin fusion proteins of the invention can be used as immunogens to produce anti-RISKMARKER antibodies in a subject, to purify RISKMARKER ligands, and in screening assays to identify molecules that inhibit the interaction of RISKMARKER with a RISKMARKER ligand.

An RISKMARKER chimeric or fusion protein of the invention can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, *e.g.*, by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers that give rise to complementary overhangs between two consecutive gene fragments that can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Ausubel *et al.* (eds.) Current Protocols in Molecular Biology, John Wiley & Sons, 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (*e.g.*, a GST polypeptide). A RISKMARKER -encoding nucleic

acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the RISKMARKER protein.

RISKMARKER AND INJURYMARKER AGONISTS AND ANTAGONISTS

The present invention also pertains to variants of the RISKMARKER or
5 INJURYMARKER proteins that function as either RISKMARKER or INJURYMARKER
agonists (mimetics) or as RISKMARKER or INJURYMARKER antagonists. Variants of the
RISKMARKER or INJURYMARKER protein can be generated by mutagenesis, *e.g.*, discrete
point mutation or truncation of the RISKMARKER or INJURYMARKER protein. An agonist
10 of the RISKMARKER or INJURYMARKER protein can retain substantially the same, or a
subset of, the biological activities of the naturally occurring form of the RISKMARKER or
INJURYMARKER protein. An antagonist of the RISKMARKER or INJURYMARKER protein
can inhibit one or more of the activities of the naturally occurring form of the RISKMARKER or
INJURYMARKER protein by, for example, competitively binding to a downstream or upstream
15 member of a cellular signaling cascade which includes the RISKMARKER or
INJURYMARKER protein. Thus, specific biological effects can be elicited by treatment with a
variant of limited function. In one embodiment, treatment of a subject with a variant having a
subset of the biological activities of the naturally occurring form of the protein has fewer side
effects in a subject relative to treatment with the naturally occurring form of the RISKMARKER
or INJURYMARKER proteins.

20 Variants of the RISKMARKER or INJURYMARKER protein that function as either
RISKMARKER or INJURYMARKER agonists (mimetics) or as RISKMARKER or
INJURYMARKER antagonists can be identified by screening combinatorial libraries of mutants,
e.g., truncation mutants, of the RISKMARKER or INJURYMARKER protein for
RISKMARKER or INJURYMARKER protein agonist or antagonist activity. In one
25 embodiment, a variegated library of RISKMARKER or INJURYMARKER variants is generated
by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene
library. A variegated library of RISKMARKER or INJURYMARKER variants can be produced
by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene
sequences such that a degenerate set of potential RISKMARKER or INJURYMARKER

sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (*e.g.*, for phage display) containing the set of RISKMARKER or INJURYMARKER sequences therein. There are a variety of methods which can be used to produce libraries of potential RISKMARKER or INJURYMARKER variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential RISKMARKER or INJURYMARKER sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, *e.g.*, Narang (1983) *Tetrahedron* 39:3; Itakura *et al.* (1984) *Annu Rev Biochem* 53:323; Itakura *et al.* (1984) *Science* 198:1056; Ike *et al.* (1983) *Nucl Acid Res* 11:477.

POLYPEPTIDE LIBRARIES

In addition, libraries of fragments of the RISKMARKER or INJURYMARKER protein coding sequence can be used to generate a variegated population of RISKMARKER or INJURYMARKER fragments for screening and subsequent selection of variants of an RISKMARKER or INJURYMARKER protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of a RISKMARKER or INJURYMARKER coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA that can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal and internal fragments of various sizes of the RISKMARKER or INJURYMARKER protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of RISKMARKER or INJURYMARKER proteins. The most widely used techniques, which are amenable to high

throughput analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected.

- 5 Recursive ensemble mutagenesis (REM), a new technique that enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify RISKMARKER or INJURYMARKER variants (Arkin and Yourvan (1992) PNAS 89:7811-7815; Delgrave *et al.* (1993) Protein Engineering 6:327-331).

ANTI-RISKMARKER AND ANTI-INJURYMARKER ANTIBODIES

- 10 An isolated RISKMARKER or INJURYMARKER protein, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind RISKMARKER or INJURYMARKER using standard techniques for polyclonal and monoclonal antibody preparation. The full-length RISKMARKER or INJURYMARKER protein can be used or, alternatively, the invention provides antigenic peptide fragments of RISKMARKER or
- 15 INJURYMARKER for use as immunogens. The antigenic peptide of RISKMARKER or INJURYMARKER comprises at least 8 amino acid residues of the amino acid sequence encoded by a nucleic acid comprising the nucleic acid sequence shown in RISKMARKER 1-8 and INJURYMARKER 1-10 and encompasses an epitope of RISKMARKER or INJURYMARKER such that an antibody raised against the peptide forms a specific immune complex with
- 20 RISKMARKER or INJURYMARKER. Preferably, the antigenic peptide comprises at least 10 amino acid residues, more preferably at least 15 amino acid residues, even more preferably at least 20 amino acid residues, and most preferably at least 30 amino acid residues. Preferred epitopes encompassed by the antigenic peptide are regions of RISKMARKER or INJURYMARKER that are located on the surface of the protein, *e.g.*, hydrophilic regions. As a
- 25 means for targeting antibody production, hydropathy plots showing regions of hydrophilicity and hydrophobicity may be generated by any method well known in the art, including, for example, the Kyte Doolittle or the Hopp Woods methods, either with or without Fourier transformation. See, *e.g.*, Hopp and Woods, 1981, Proc. Nat. Acad. Sci. USA 78: 3824-3828; Kyte and Doolittle 1982, J. Mol. Biol. 157: 105-142, each incorporated herein by reference in their entirety.

RISKMARKER or INJURYMARKER polypeptides or derivatives, fragments, analogs or homologs thereof, may be utilized as immunogens in the generation of antibodies that immunospecifically-bind these protein components. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen binding site that specifically binds (immunoreacts with) an antigen. Such antibodies include, but are not limited to, polyclonal, monoclonal, chimeric, single chain, F_{ab} and $F_{(ab)2}$ fragments, and an F_{ab} expression library. Various procedures known within the art may be used for the production of polyclonal or monoclonal antibodies to an RISKMARKER or INJURYMARKER protein sequence, *e.g.* RISKMAKER 1 or RISKMAKER 6-8, or derivatives, fragments, analogs or homologs thereof. Some of these proteins are discussed below.

For the production of polyclonal antibodies, various suitable host animals (*e.g.*, rabbit, goat, mouse or other mammal) may be immunized by injection with the native protein, or a synthetic variant thereof, or a derivative of the foregoing. An appropriate immunogenic preparation can contain, for example, recombinantly expressed RISKMARKER or INJURYMARKER protein or a chemically synthesized RISKMARKER or INJURYMARKER polypeptide. The preparation can further include an adjuvant. Various adjuvants used to increase the immunological response include, but are not limited to, Freund's (complete and incomplete), mineral gels (*e.g.*, aluminum hydroxide), surface active substances (*e.g.*, lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, dinitrophenol, etc.), human adjuvants such as *Bacille Calmette-Guerin* and *Corynebacterium parvum*, or similar immunostimulatory agents. If desired, the antibody molecules directed against RISKMARKER or INJURYMARKER can be isolated from the mammal (*e.g.*, from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG fraction.

The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of RISKMARKER or INJURYMARKER. A monoclonal antibody composition thus typically displays a single binding affinity for a particular RISKMARKER or INJURYMARKER protein with which it immunoreacts. For preparation of monoclonal antibodies directed towards a particular RISKMARKER or INJURYMARKER protein, or derivatives, fragments, analogs or homologs

thereof, any technique that provides for the production of antibody molecules by continuous cell line culture may be utilized. Such techniques include, but are not limited to, the hybridoma technique (see Kohler & Milstein, 1975 *Nature* 256: 495-497); the trioma technique; the human B-cell hybridoma technique (see Kozbor, *et al.*, 1983 *Immunol Today* 4: 72) and the EBV

hybridoma technique to produce human monoclonal antibodies (see Cole, *et al.*, 1985 In: Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). Human monoclonal antibodies may be utilized in the practice of the present invention and may be produced by using human hybridomas (see Cote, *et al.*, 1983. *Proc Natl Acad Sci USA* 80: 2026-2030) or by transforming human B-cells with Epstein Barr Virus *in vitro* (see Cole, *et al.*, 1985 In: Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96).

According to the invention, techniques can be adapted for the production of single-chain antibodies specific to a RISKMARKER or INJURYMARKER protein (see *e.g.*, U.S. Patent No. 4,946,778). In addition, methods can be adapted for the construction of F_{ab} expression libraries (see *e.g.*, Huse, *et al.*, 1989 *Science* 246: 1275-1281) to allow rapid and effective identification of monoclonal F_{ab} fragments with the desired specificity for a RISKMARKER or INJURYMARKER protein or derivatives, fragments, analogs or homologs thereof. Non-human antibodies can be "humanized" by techniques well known in the art. See *e.g.*, U.S. Patent No. 5,225,539. Antibody fragments that contain the idiotypes to a RISKMARKER or INJURYMARKER protein may be produced by techniques known in the art including, but not limited to: (i) an F_{(ab)2} fragment produced by pepsin digestion of an antibody molecule; (ii) an F_{ab} fragment generated by reducing the disulfide bridges of an F_{(ab)2} fragment; (iii) an F_{ab} fragment generated by the treatment of the antibody molecule with papain and a reducing agent and (iv) F_v fragments.

Additionally, recombinant anti-RISKMARKER or INJURYMARKER antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in PCT International Application No. PCT/US86/02269; European Patent Application No. 184,187; European Patent Application No. 171,496; European Patent Application No. 173,494; PCT International Publication No. WO 86/01533; U.S. Pat. No. 4,816,567; European Patent

Application No. 125,023; Better *et al.* (1988) *Science* 240:1041-1043; Liu *et al.* (1987) *PNAS* 84:3439-3443; Liu *et al.* (1987) *J Immunol.* 139:3521-3526; Sun *et al.* (1987) *PNAS* 84:214-218; Nishimura *et al.* (1987) *Cancer Res* 47:999-1005; Wood *et al.* (1985) *Nature* 314:446-449; Shaw *et al.* (1988) *J Natl Cancer Inst.* 80:1553-1559; Morrison (1985) *Science* 229:1202-1207; Oi *et al.* (1986) *BioTechniques* 4:214; U.S. Pat. No. 5,225,539; Jones *et al.* (1986) *Nature* 321:552-525; Verhoeyan *et al.* (1988) *Science* 239:1534; and Beidler *et al.* (1988) *J Immunol* 141:4053-4060.

In one embodiment, methods for the screening of antibodies that possess the desired specificity include, but are not limited to, enzyme-linked immunosorbent assay (ELISA) and other immunologically-mediated techniques known within the art. In a specific embodiment, selection of antibodies that are specific to a particular domain of a RISKMARKER or INJURYMARKER protein is facilitated by generation of hybridomas that bind to the fragment of a RISKMARKER or INJURYMARKER protein possessing such a domain. Antibodies that are specific for one or more domains within a RISKMARKER or INJURYMARKER protein, *e.g.*, domains spanning the above-identified conserved regions of RISKMARKER or INJURYMARKER family proteins, or derivatives, fragments, analogs or homologs thereof, are also provided herein.

Anti-RISKMARKER or anti-INJURYMARKER antibodies may be used in methods known within the art relating to the localization and/or quantitation of a RISKMARKER or INJURYMARKER protein (*e.g.*, for use in measuring levels of the RISKMARKER or INJURYMARKER protein within appropriate physiological samples, for use in diagnostic methods, for use in imaging the protein, and the like). In a given embodiment, antibodies for RISKMARKER or INJURYMARKER proteins, or derivatives, fragments, analogs or homologs thereof, that contain the antibody derived binding domain, are utilized as pharmacologically-active compounds [hereinafter "Therapeutics"].

An anti-RISKMARKER or INJURYMARKER antibody (*e.g.*, monoclonal antibody) can be used to isolate RISKMARKER or INJURYMARKER by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-RISKMARKER or INJURYMARKER antibody can facilitate the purification of natural RISKMARKER or INJURYMARKER from cells and of recombinantly produced RISKMARKER or INJURYMARKER expressed in host

cells. Moreover, an anti-RISKMARKER or INJURYMARKER antibody can be used to detect RISKMARKER or INJURYMARKER protein (*e.g.*, in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the RISKMARKER or INJURYMARKER protein. Anti-RISKMARKER or INJURYMARKER antibodies can be used

5 diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, *e.g.*, to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling (*i.e.*, physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable

10 enzymes include horseradish peroxidase, alkaline phosphatase, β -galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of

15 bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S or ^3H .

RISKMARKER RECOMBINANT VECTORS AND HOST CELLS

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding RISKMARKER protein, *e.g.*, RISKMARKER 1, or

20 RISKMARKER 6-8, or derivatives, fragments, analogs or homologs thereof. As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a linear or circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome.

25 Certain vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes

30 to which they are operatively linked. Such vectors are referred to herein as "expression vectors".

In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, that is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner that allows for expression of the nucleotide sequence (*e.g.*, in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (*e.g.*, polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, Calif. (1990). Regulatory sequences include those that direct constitutive expression of a nucleotide sequence in many types of host cell and those that direct expression of the nucleotide sequence only in certain host cells (*e.g.*, tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (*e.g.*, RISKMARKER proteins, mutant forms, fusion proteins, etc.).

The recombinant expression vectors of the invention can be designed for expression of RISKMARKER in prokaryotic or eukaryotic cells. For example, RISKMARKER can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, Calif. (1990). Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: (1) to increase expression of recombinant protein; (2) to increase the solubility of the recombinant protein; and (3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith and Johnson (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, Mass.) and pRIT5 (Pharmacia, Piscataway, N.J.) that fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amrann *et al.*, (1988) *Gene* 69:301-315) and pET 11d (Studier *et al.*, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, Calif. (1990) 60-89).

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein. See, Gottesman, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, Calif. (1990) 119-128. Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada *et al.*, (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the RISKMARKER expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari, *et al.*, (1987) *EMBO J* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.*, (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, Calif.), and picZ (Invitrogen Corp, San Diego, Calif.).

Alternatively, RISKMARKER can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (*e.g.*, SF9 cells) include the pAc series (Smith *et al.* (1983) *Mol Cell Biol* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

5 In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed (1987) *Nature* 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J* 6: 187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from
10 polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells. See, *e.g.*, Chapters 16 and 17 of Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*. 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989.

15 In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (*e.g.*, tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert *et al.* (1987) *Genes Dev* 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) *Adv Immunol* 43:235-275),
20 in particular promoters of T cell receptors (Winoto and Baltimore (1989) *EMBO J* 8:729-733) and immunoglobulins (Banerji *et al.* (1983) *Cell* 33:729-740; Queen and Baltimore (1983) *Cell* 33:741-748), neuron-specific promoters (*e.g.*, the neurofilament promoter; Byrne and Ruddle (1989) *PNAS* 86:5473-5477), pancreas-specific promoters (Edlund *et al.* (1985) *Science* 230:912-916), and mammary gland-specific promoters (*e.g.*, milk whey promoter; U.S. Pat. No.
25 4,873,316 and European Application Publication No. 264,166). Developmentally-regulated promoters are also encompassed, *e.g.*, the murine hox promoters (Kessel and Gruss (1990) *Science* 249:374-379) and the α -fetoprotein promoter (Campes and Tilghman (1989) *Genes Dev* 3:537-546).

The invention further provides a recombinant expression vector comprising a DNA
30 molecule of the invention cloned into the expression vector in an antisense orientation. That is,

the DNA molecule is operatively linked to a regulatory sequence in a manner that allows for expression (by transcription of the DNA molecule) of an RNA molecule that is antisense to RISKMARKER mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen that direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen that direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub *et al.*, "Antisense RNA as a molecular tool for genetic analysis," Reviews--Trends in Genetics, Vol. 1(1) 1986.

Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but also to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, RISKMARKER protein can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, *et al.*

(Molecular Cloning: A Laboratory Manual. 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (*e.g.*, resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Various selectable markers include those that confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding RISKMARKER or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) an RISKMARKER protein, *e.g.* RISKMARKER 1, or RISKMARKER 6-8. Accordingly, the invention further provides methods for producing RISKMARKER protein using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding RISKMARKER has been introduced) in a suitable medium such that RISKMARKER protein is produced. In another embodiment, the method further comprises isolating RISKMARKER from the medium or the host cell.

PHARMACEUTICAL COMPOSITIONS

The RISKMARKER nucleic acid molecules, RISKMARKER proteins, and anti-RISKMARKER or anti-INJURYMARKER antibodies (also referred to herein as "active compounds") of the invention, and derivatives, fragments, analogs and homologs thereof, can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically acceptable carrier. As used herein, "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and

absorption delaying agents, and the like, compatible with pharmaceutical administration. Suitable carriers are described in the most recent edition of Remington's Pharmaceutical Sciences, a standard reference text in the field, which is incorporated herein by reference. Preferred examples of such carriers or diluents include, but are not limited to, water, saline, finger's solutions, dextrose solution, and 5% human serum albumin. Liposomes and non-aqueous vehicles such as fixed oils may also be used. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, *e.g.*, intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates, and agents for the adjustment of tonicity such as sodium chloride or dextrose. The pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor EL™ (BASF, Parsippany, N.J.) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringeability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol,

and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a RISKMARKER protein or anti-RISKMARKER or INJURYMARKER antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle that contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, methods of preparation are vacuum drying and freeze-drying that yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, *e.g.*, a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For
5 transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into
10 ointments, salves, gels, or creams as generally known in the art. The compounds can also be prepared in the form of suppositories (*e.g.*, with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation,
15 including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes
20 targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Pat. No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers
25 to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and

the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Pat. No. 5,328,470) or by stereotactic injection (see *e.g.*, Chen *et al.* (1994) *PNAS* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, *e.g.*, retroviral vectors, the pharmaceutical preparation can include one or more cells that produce the gene delivery system. The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

KITS AND NUCLEIC ACID COLLECTIONS FOR IDENTIFYING RISKMARKER AND INJURYMARKER NUCLEIC ACIDS

In another aspect, the invention provides a kit useful for examining hepatotoxicity of agents. The kit can include nucleic acids that detect two or more RISKMARKER or INJURYMARKER sequences. In preferred embodiments, the kit includes reagents which detect 3, 4, 5, 6, 8, 10, 12, 15, or all of the RISKMARKER or INJURYMARKER nucleic acid sequences.

The invention also includes an isolated plurality of sequences which can identify one or more RISKMARKER or INJURYMARKER responsive nucleic acid sequences. The kit or plurality may include, *e.g.*, sequence homologous to RISKMARKER or INJURYMARKER nucleic acid sequences, or sequences which can specifically identify one or more RISKMARKER or INJURYMARKER nucleic acid sequences.

OTHER EMBODIMENTS

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the

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WE CLAIM:

1. A method of screening a test agent for hepatotoxicity, the method comprising;
 - (a) providing a test cell population comprising a cell capable of expressing one or more nucleic acid sequences selected from the group consisting of RISKMARKER 1-8 and INJURYMARKER 1-10;
 - (b) contacting the test cell population with a test agent;
 - (c) measuring expression of one or more of the nucleic acid sequences in the test cell population;
 - (d) comparing the expression of the nucleic acid sequences in the test cell population to the expression of the nucleic acid sequences in a reference cell population comprising at least one cell whose exposure status to a hepatotoxic agent is known; and
 - (e) identifying a difference in expression levels of the RISKMARKER or INJURYMARKER sequences, if present, in the test cell population and reference cell population,thereby screening said test agent for hepatotoxicity.
2. The method of claim 1, wherein said hepatotoxicity comprises idiosyncratic hepatotoxicity.
3. The method of claim 2, wherein the method comprises comparing the expression of one or more nucleic acid sequences selected from the group consisting of RISKMARKER 1-8.
4. The method of claim 2, wherein the method comprises comparing the expression of one or more nucleic acid sequences selected from the group consisting of INJURYMARKER 1-10.
5. The method of claim 1, wherein the method comprises comparing the expression of 6 or more of the nucleic acid sequences.

6. The method of claim 1, wherein the expression of the nucleic acid sequences in the test cell population is decreased as compared to the reference cell population.
7. The method of claim 1, wherein the expression of the nucleic acid sequences in the test cell population is increased as compared to the reference cell population.
8. The method of claim 1, wherein the test cell population is provided in vitro.
9. The method of claim 1, wherein the test cell population is provided ex vivo from a mammalian subject.
10. The method of claim 1, wherein the test cell population is provided in vivo in a mammalian subject.
11. The method of claim 1, wherein the test cell population is derived from a human or rodent subject.
12. The method of claim 1, wherein the test cell population includes a hepatocyte.
13. The method of claim 1, wherein said test agent is an idiosyncratic hepatotoxic agent.
14. The method of claim 1, wherein said test agent is a non-steriodal anti-inflammatory drug (NSAID).
15. The method of claim 3, wherein said hepatotoxic agent is a NSAID.
16. The method of claim 15, wherein said NSAID is a NSAID classified as having a low risk of hepatotoxicity, and wherein said test agent is identified as having a low risk of hepatotoxicity if no qualitative difference in expression levels is identified in step (e).

17. The method of claim 16, wherein said difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations.
18. The method of claim 16, wherein said NSAID is selected from the group consisting of Benoxaprofen, Bromfenac, Diclofenac, Phenylbutazone, and Sulindac.
19. The method of claim 18, wherein said NSAID is selected from the group consisting of Benoxaprofen, Phenylbutazone, and Sulindac.
20. The method of claim 15, wherein said NSAID is a NSAID classified as having a very low risk of hepatotoxicity, and wherein said test agent is identified as having a very low risk of hepatotoxicity if no qualitative difference in expression levels is identified in step (e).
21. The method of claim 20, wherein said difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations.
22. The method of claim 20, wherein said NSAID is selected from the group consisting of Etodolac, Fenoprofen, Flurbiprofen, Ibuprofen, Indomethacin, Ketoprofen, Meclofenamate, Mefenamic Acid, Nabumetone, Naproxen, Oxaprozin, Piroxicam, Suprofen, Tenoxicam, Tolmentin, and Zomepirac.
23. The method of claim 22, wherein said NSAID is selected from the group consisting of Flurbiprofen, Oxaprozin, and Tenoxicam.
24. The method of claim 15, wherein said NSAID is a NSAID classified as having an overdose risk of hepatotoxicity, and wherein said test agent is identified as having an overdose risk of hepatotoxicity if no qualitative difference in expression levels is identified in step (e).

25. The method of claim 24, wherein said difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations.
26. The method of claim 25, wherein said NSAID is selected from the group consisting of Acetaminophen, Acetylsalicylic acid, and Phenacetin.
27. The method of claim 4, wherein said hepatotoxic agent is a NSAID.
28. The method of claim 27, wherein said NSAID is a NSAID classified as inducing hepatocellular damage, and wherein said test agent is identified as likely to induce hepatocellular damage if no qualitative difference in expression levels is identified in step (e).
29. The method of claim 28, wherein said difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations.
30. The method of claim 27, wherein said NSAID is selected from the group consisting of Acetaminophen, Flurbiprofen, and Ketoprofen.
31. The method of claim 27, wherein said NSAID is a NSAID classified as inducing cholestasis, and wherein said test agent is identified as likely to induce cholestasis if no qualitative difference in expression levels is identified in step (e).
32. The method of claim 31, wherein said difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations.

- 33 The method of claim 30, wherein said NSAID is selected from the group consisting of Benoxaprofen, Nabumetone, and Sulindac.
34. The method of claim 27, wherein said NSAID is a NSAID classified as inducing elevated transaminase level, and wherein said test agent is identified as likely to induce elevated transaminase level if no qualitative difference in expression levels is identified in step (e).
35. The method of claim 34, wherein said difference in expression levels is determined by comparing expression transformation eigenvectors for said test cell and reference cell populations.
36. The method of claim 34, wherein said NSAID is selected from the group consisting of Zomepirac, Mefenamic acid, and Tenoxicam.
37. A method of assessing the hepatotoxicity of a test agent in a subject, the method comprising:
- (a) providing from the subject a test cell population comprising a cell capable of expressing one or more nucleic acid sequences selected from the group consisting of RISKMARKER 1-8 and INJURYMARKER 1-10;
 - (b) contacting the test cell population with a test agent;
 - (c) measuring expression of one or more of the nucleic acid sequences in the test cell population; and
 - (d) comparing the expression of the nucleic acid sequences in the test cell population to the expression of the nucleic acid sequences in a reference cell population comprising at least one cell whose exposure status to a hepatotoxic agent is known;
 - (e) identifying a difference in expression levels of the nucleic acid sequences, if present, in the test cell population and the reference cell population, thereby assessing the hepatotoxicity of the test agent in the subject.

38. The method of claim 37, wherein said hepatotoxicity comprises idiosyncratic hepatotoxicity.

39. The method of claim 38, wherein the method comprises comparing the expression of one or more nucleic acid sequences selected from the group consisting of RISKMARKER 1-8.

40. The method of claim 38, wherein the method comprises comparing the expression of one or more nucleic acid sequences selected from the group consisting of INJURYMARKER 1-10.

41. The method of claim 37, wherein the expression of the nucleic acid sequences in the test cell population is increased as compared to the reference cell population.

42. The method of claim 37, wherein the expression of the nucleic acid sequences in the test cell population is increased as compared to the reference cell population.

43. The method of claim 37, wherein said subject is a human or rodent.

44. The method of claim 37, wherein the test cell population is provided ex vivo from said subject.

45. The method of claim 37, wherein the test cell population is provided in vivo from said subject.

46. The method of claim 37, wherein said test agent is a non-steroidal anti-inflammatory drug (NSAID).

47. The method of claim 37, wherein said hepatotoxic agent is a NSAID.

48. An isolated nucleic acid comprising a nucleic acid sequence selected from the group consisting of a RISKMARKER 1 nucleic acid, a RISKMARKER 6-8 nucleic acid, and their complements.

49. A vector comprising the nucleic acid of claim 48.

50. A cell comprising the vector of claim 49.

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51. A pharmaceutical composition comprising the nucleic acid of claim 48.

52. A polypeptide encoded by the nucleic acid of claim 48.

10 53. An antibody which specifically binds to the polypeptide of claim 52.

54. A kit which detects two or more of the nucleic acid sequences selected from the group consisting of RISKMARKER 1, and RISKMARKER 6-8.

15 55. An array which detects one or more of the nucleic acid selected from the group consisting of RISKMARKER 1, and RISKMARKER 6-8.

56. A plurality of nucleic acid comprising one or more of the nucleic acid selected from the group consisting of RISKMARKER 1, and RISKMARKER 6-8.

**METHOD OF IDENTIFYING TOXIC AGENTS USING NSAID-INDUCED
DIFFERENTIAL GENE EXPRESSION IN LIVER**

ABSTRACT

The invention provides methods of identifying toxic agents, *e.g.*, hepatotoxic agents,
5 using differential gene expression. Also provided are methods of predicting the risk level and or
injury type of NSAIDs. Also disclosed are novel nucleic acid sequences whose expression is
differentially regulated by NSAIDs.

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COMBINED DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am an original, first and sole inventor of the subject matter (an original, first and joint inventor) which is claimed and for which a utility patent is sought on the invention entitled:

METHOD OF IDENTIFYING TOXIC AGENTS USING NSAID-INDUCED DIFFERENTIAL GENE EXPRESSION IN LIVER

the specification of which:

- ☐ was filed on _____, as United States non-provisional application
U.S.S.N. _____, bearing Attorney Docket No. _____.
- ☒ is attached hereto.

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56.

- ☐ I hereby claim foreign priority benefits under Title 35, United States Code, §119(a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate, or §365(a) of any PCT International application designating at least one country other than the United States listed below and have also identified below any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed.

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- ☒ I hereby claim the benefit under Title 35, United States Code, § 119(e) or §120 of any United States application(s), or §365(c) of any PCT International application(s) designating the United States of America listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

Application No. (U.S.S.N.)	Filing Date (dd/mm/yy)	Status (Patented, Pending, Abandoned)
60/166,923	November 22, 1999	Pending
60/183,531	February 18, 2000	Pending

PCT International Applications designating the United States:

PCT International Application No.	PCT Filing Date	Status

I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

Attorney or Agent	Registration No.	Attorney or Agent	Registration No.
Kevin Ainsworth	39,586	Kristin E. Konzak	44,848
Ingrid Beattie	42,306	Cynthia Kozakiewicz	42,764
David B. Bernstein	32,112	Barry Marenberg	40,715
Naomi Biswas	38,384	William Marino	44,219
David F. Crosby	36,400	A. Jason Mirabito	28,161
James G. Cullem	43,569	Michel Morency	Limited Recognition
Brett N. Dorny	35,860	Bradley Olson	40,750
Marianne Downing	42,870	Carol H. Peters	45,010
Ivor R. Elrifi	39,529	John T. Prince	43,091
Heidi A. Erlacher	45,409	Michael Renaud	44,299
Christina Gadiano	37,628	Brian Rosenbloom	41,276
John A. Harre	37,345	Thomas M. Sullivan	39,392
Shane Hunter	41,858	Howard Susser	33,556
David E. Johnson	41,874	Shelby J. Walker	45,192
Kris Kalidindi	41,461	Martin M. Zoltick	35,745
Christina Karnakis	45,899		
Robert Klauzinski	42,742		

all of MINTZ, LEVIN, COHN, FERRIS, GLOVSKY AND POPEO PC, One Financial Center, Boston, Massachusetts 02111, as Applicant's attorneys with full power of substitution and revocation to take any and all action necessary with regard to the above-identified patent.

Address all telephone calls to Ivor R. Elrifi, Esq., at telephone number 617/542-6000.
Address all correspondence to:

Ivor R. Elrifi, Esq.
Mintz, Levin, Cohn, Ferris, Glovsky and Popeo, P.C.
One Financial Center
Boston, Massachusetts 02111

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or patent issued thereon.

Inventor's Signature
Full Name of Inventor: Bonnie Gould Rothberg
Citizenship: USA
Residence: 1701 Moose Hill Road
Post Office Address: Guilford, CT 06437

Date

Inventor's Signature
Full Name of Inventor: Vincent A. DiPippio
Citizenship: USA
Residence: 156 North Atwater Street
Post Office Address: East Haven, CT 06512

Date

Inventor's Signature
Full Name of Inventor: Tenmore M. Ramesh
Citizenship: India
Residence: 7 Sandpiper Lane
Post Office Address: New Milford, CT 06776

Date

Date _____

Citizenship: USA

Post Office Address: Brandford, CT 06405

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